

# Package ‘scan’

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**Type** Package

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**Description** A collection of procedures for analysing, visualising, and managing single-case data. These include piecewise linear regression models, multilevel models, overlap indices (PND, PEM, PAND, PET, tau-u, baseline corrected tau, CDC), and randomization tests. Data preparation functions support outlier detection, handling missing values, scaling, truncating, rank transformation, and smoothing. An exporting function helps to generate html and latex tables in a publication friendly style. More details can be found at <<https://jazznbass.github.io/scan-Book/>>.

**License** GPL (>= 3)

**URL** <https://github.com/jazznbass/scan/>,  
<https://jazznbass.github.io/scan-Book/>

**BugReports** <https://github.com/jazznbass/scan/issues/>

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<code>add_l2</code>	<i>Add level-2 data</i>
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---

**Description**

Merges variables with corresponding case names from a data.frame with an scdf file

**Usage**

```
add_l2(scdf, data_l2, cvar = "case")
```

**Arguments**

- `scdf`            A single-case data frame. See [scdf](#) to learn about this format.
- `data_l2`        A level 2 dataset.
- `cvar`            Character string with the name of the "case" variable in the L2 dataset (default is 'case').

**Details**

This function is mostly used in combination with the [hplm](#) function.

**Value**

An scdf

**See Also**

- [hplm](#)
- Other data manipulation functions: [as.data.frame.scdf\(\)](#), [as.scdf\(\)](#), [fill\\_missing\(\)](#), [moving\\_median\(\)](#), [outlier\(\)](#), [ranks\(\)](#), [scdf](#), [select\\_cases\(\)](#), [set\\_vars\(\)](#), [shift\(\)](#), [smooth\\_cases\(\)](#), [standardize\(\)](#), [truncate\\_phase\(\)](#)

**Examples**

```
Leidig2018 %>% add_l2(Leidig2018_l2)
```

---

as.data.frame.scdf      *Creating a long format data frame from several single-case data frames (scdf).*

---

### Description

The `as.data.frame` function transposes an `scdf` into one long data frame. Additionally, a data frame can be merged that includes level 2 data of the subjects. This might be helpful to prepare data to be used with other packages than `scan`.

### Usage

```
## S3 method for class 'scdf'  
as.data.frame(x, ..., l2 = NULL, id = "case")
```

### Arguments

<code>x</code>	An <code>scdf</code> object
<code>...</code>	Not implemented
<code>l2</code>	A data frame providing additional variables at Level 2. The <code>scdf</code> has to have names for all cases and the Level 2 data frame has to have a column with corresponding case names.
<code>id</code>	Variable name of the Level 2 data frame that contains the case names.

### Value

Returns one data frame with data of all single-cases structured by the case variable.

### Author(s)

Juergen Wilbert

### See Also

Other data manipulation functions: [add\\_l2\(\)](#), [as\\_scdf\(\)](#), [fill\\_missing\(\)](#), [moving\\_median\(\)](#), [outlier\(\)](#), [ranks\(\)](#), [scdf](#), [select\\_cases\(\)](#), [set\\_vars\(\)](#), [shift\(\)](#), [smooth\\_cases\(\)](#), [standardize\(\)](#), [truncate\\_phase\(\)](#)

### Examples

```
## Convert the list of three single-case data frames from Grosche (2011)  
### into one long data frame  
Grosche2011  
Grosche2011_long <- as.data.frame(Grosche2011)  
Grosche2011_long
```

```
## Combine an scdf with data for l2
Leidig2018_long <- as.data.frame(Leidig2018, l2 = Leidig2018_l2)
names(Leidig2018_long)
summary(Leidig2018_long)
```

---

as\_scdf

*as\_scdf*

---

## Description

Converts a data frame to an scdf object.

## Usage

```
as_scdf(  
  object,  
  cvar = "case",  
  pvar = "phase",  
  dvar = "values",  
  mvar = "mt",  
  phase_names = NULL,  
  sort_cases = FALSE  
)
```

## Arguments

object	A data.frame
cvar	Sets the variable name of the "case" variable. Defaults to "case".
pvar	Sets the variable name of the "phase" variable. Defaults to "phase".
dvar	Sets the variable name of the "values" variable. Defaults to "values".
mvar	Sets the variable name of the "mt" variable. Defaults to "mt".
phase_names	A character vector with phase names. Defaults to the phase names provided in the phase variable.
sort_cases	If set TRUE, the resulting list is sorted by label names (alphabetically increasing).

## Value

An scdf.

## See Also

Other data manipulation functions: [add\\_l2\(\)](#), [as.data.frame.scdf\(\)](#), [fill\\_missing\(\)](#), [moving\\_median\(\)](#), [outlier\(\)](#), [ranks\(\)](#), [scdf](#), [select\\_cases\(\)](#), [set\\_vars\(\)](#), [shift\(\)](#), [smooth\\_cases\(\)](#), [standardize\(\)](#), [truncate\\_phase\(\)](#)

---

`autocorr`*Autocorrelation for single-case data*

---

**Description**

The autocorr function calculates autocorrelations within each phase and across all phases.

**Usage**

```
autocorr(data, dvar, pvar, mvar, lag_max = 3, lag.max, ...)
```

**Arguments**

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
<code>mvar</code>	Character string with the name of the measurement time variable. Defaults to the attributes in the scdf file.
<code>lag_max, lag.max</code>	The lag up to which autocorrelations will be computed.
<code>...</code>	Further arguments passed to the <a href="#">acf</a> function

**Value**

A data frame containing separate autocorrelations for each phase and for all phases (for each single-case). If `lag_max` exceeds the length of a phase minus one, NA is returned for this cell.

**Author(s)**

Juergen Wilbert

**See Also**

[acf](#)

Other regression functions: [corrected\\_tau\(\)](#), [hplm\(\)](#), [mplm\(\)](#), [plm\(\)](#), [trend\(\)](#)

**Examples**

```
## Compute autocorrelations for a list of four single-cases up to lag 2.
autocorr(Huber2014, lag_max = 2)
```

**Description**

The `cdc` function applies the Conservative Dual-Criterion Method (Fisher, Kelley, & Lomas, 2003) to `scdf` objects. It compares phase B data points to both phase A mean and trend (OLS, bi-split, tri-split) with an additional increase/decrease of .25 SD. A binomial test against a 50/50 distribution is computed and p-values below .05 are labeled "systematic change".

**Usage**

```
cdc(
  data,
  dvar,
  pvar,
  mvar,
  decreasing = FALSE,
  trend_method = c("OLS", "bisplit", "trisplit"),
  conservative = 0.25,
  phases = c(1, 2)
)
```

**Arguments**

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the <code>scdf</code> file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the <code>scdf</code> file.
<code>mvar</code>	Character string with the name of the measurement time variable. Defaults to the attributes in the <code>scdf</code> file.
<code>decreasing</code>	If you expect data to be lower in the B phase, set <code>decreasing = TRUE</code> . Default is <code>decreasing = FALSE</code> .
<code>trend_method</code>	Method used to calculate the trend line. Default is <code>trend_method = "OLS"</code> . Possible values are: "OLS", "bisplit", and "trisplit". "bisplit", and "trisplit" should only be used for cases with at least five data-points in both relevant phases.
<code>conservative</code>	The CDC method adjusts the original mean and trend lines by adding (expected increase) or subtracting (expected decrease) an additional .25 SD before evaluating phase B data. Default is the CDC method with <code>conservative = .25</code> . To apply the Dual-Criterion (DC) method, set <code>conservative = 0</code> .
<code>phases</code>	A vector of two characters or numbers indicating the two phases that should be compared. E.g., <code>phases = c("A", "C")</code> or <code>phases = c(2, 4)</code> for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., <code>phases = list(A = c(1, 3), B = c(2, 4))</code> will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is <code>phases = c(1, 2)</code> .

**Value**

cdc	CDC Evaluation based on a p-value below .05.
cdc_exc	Number of phase B datapoints indicating expected change.
cdc_nb	Number of phase B datapoints.
cdc_p	P value of Binomial Test.
cdc_all	Overall CDC Evaluation based on all instances/cases of a Multiple Baseline Design.
N	Number of cases.
decreasing	Logical argument from function call (see Arguments above).
conservative	Numeric argument from function call (see Arguments above).
case_names	Assigned name of single-case.
phases	-

**Author(s)**

Timo Lueke

**References**

Fisher, W. W., Kelley, M. E., & Lomas, J. E. (2003). Visual Aids and Structured Criteria for Improving Visual Inspection and Interpretation of Single-Case Designs. *Journal of Applied Behavior Analysis*, 36, 387-406. <https://doi.org/10.1901/jaba.2003.36-387>

**Examples**

```
## Apply the CDC method (standard OLS line)
design <- design(n = 1, slope = 0.2)
dat <- random_scdf(design, seed = 42)
cdc(dat)

## Apply the CDC with Koenig's bi-split and an expected decrease in phase B.
cdc(exampleAB_decreasing, decreasing = TRUE, trend_method = "bisplit")

## Apply the CDC with Tukey's tri-split, comparing the first and fourth phase.
cdc(exampleABAB, trend_method = "trisplit", phases = c(1,4))

## Apply the Dual-Criterion (DC) method (i.e., mean and trend without shifting).
cdc(exampleAB_decreasing, decreasing = TRUE, trend_method = "bisplit", conservative = 0)
```

---

coef.sc_plm	<i>Extract coefficients from plm/hplm objects</i>
-------------	---------------------------------------------------

---

**Description**

Extract coefficients from plm/hplm objects

**Usage**

```
## S3 method for class 'sc_plm'  
coef(object, ...)  
  
## S3 method for class 'sc_hplm'  
coef(object, ...)
```

**Arguments**

object	plm or hplm object
...	not implemented

**Value**

data frame with coefficient table

**Examples**

```
coefficients(plm(exampleAB$Johanna))
```

---

combine	<i>Combine single-case data frames</i>
---------	----------------------------------------

---

**Description**

Combine single-case data frames

**Usage**

```
combine(..., dvar = NULL, pvar = NULL, mvar = NULL)  
  
## S3 method for class 'scdf'  
c(...)
```

**Arguments**

...	scdf objects
dvar	Character string. Name of the dependent variable. Defaults to the dependent variable of the first case provided.
pvar	Character string. Name of the phase variable. Defaults to the phase variable of the first case provided.
mvar	Character string. Name of the measurement-time variable. Defaults to the measurement-time variable of the first case provided.

**Value**

A scdf. If not set differently, the attributes of this scdf are copied from the first scdf provided (i.e the first argument of the function).

---

convert	<i>Convert Converts an scdf object into R code</i>
---------	----------------------------------------------------

---

**Description**

Convert Converts an scdf object into R code

**Usage**

```
convert(scdf, file = "", study_name = "study", case_name = "case", indent = 3)
```

**Arguments**

scdf	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
file	A filename for exporting the syntax.
study_name	Character string. Name of the study object.
case_name	Character string. Name of the scdf objects.
indent	Integer. Indentation.

**See Also**

Other io-functions: [read\\_scdf\(\)](#), [write\\_scdf\(\)](#)

**Examples**

```
filename <- tempfile()
convert(exampleABC, file = filename)
source(filename)
all.equal(study, exampleABC)
unlink(filename)
```

---

corrected_tau	<i>Baseline corrected tau</i>
---------------	-------------------------------

---

### Description

Kendall's tau correlation for the dependent variable and the phase variable is calculated after correcting for a baseline trend.

### Usage

```
corrected_tau(
  data,
  dvar,
  pvar,
  mvar,
  phases = c(1, 2),
  alpha = 0.05,
  continuity = FALSE,
  repeated = FALSE
)
```

### Arguments

data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
dvar	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
pvar	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
mvar	Character string with the name of the measurement time variable. Defaults to the attributes in the scdf file.
phases	A vector of two characters or numbers indicating the two phases that should be compared. E.g., <code>phases = c("A", "C")</code> or <code>phases = c(2, 4)</code> for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., <code>phases = list(A = c(1, 3), B = c(2, 4))</code> will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is <code>phases = c(1, 2)</code> .
alpha	Sets the p-value at and below which a baseline correction is applied.
continuity	If TRUE applies a continuity correction for calculating p
repeated	If TRUE applies the repeated median method for calculating slope and intercept ( <a href="#">mb1m</a> )

### Details

This method has been proposed by Tarlow (2016). The baseline data are checked for a significant autocorrelation (based on Kendall's Tau). If so, a non-parametric Theil-Sen regression is applied for the baseline data where the dependent values are regressed on the measurement time. The

resulting slope information is then used to predict data of the B-phase. The dependent variable is now corrected for this baseline trend and the residuals of the Theil-Sen regression are taken for further calculations. Finally, Kendall's tau is calculated for the dependent variable and the dichotomous phase variable. The function here provides two extensions to this procedure: The more accurate Siegel repeated median regression is applied when `repeated = TRUE` and a continuity correction is applied when `continuity = TRUE`.

## References

Tarlow, K. R. (2016). An Improved Rank Correlation Effect Size Statistic for Single-Case Designs: Baseline Corrected Tau. *Behavior Modification*, 41(4), 427–467. <https://doi.org/10.1177/0145445516676750>

## See Also

Other regression functions: `autocorr()`, `hplm()`, `mplm()`, `plm()`, `trend()`

Other overlap functions: `nap()`, `overlap()`, `pand()`, `pem()`, `pet()`, `pnd()`, `tau_u()`

## Examples

```
dat <- scdf(c(A = 33,25,17,25,14,13,15, B = 15,16,16,5,7,9,6,5,3,3,8,11,7))
corrected_tau(dat)
```

---

describe

*Descriptive statistics for single-case data*

---

## Description

The `describe` function provides common descriptive statistics for single-case data.

## Usage

```
describe(data, dvar, pvar, mvar)
```

## Arguments

<code>data</code>	A single-case data frame. See <code>scdf</code> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the <code>scdf</code> file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the <code>scdf</code> file.
<code>mvar</code>	Character string with the name of the measurement time variable. Defaults to the attributes in the <code>scdf</code> file.

## Details

`n` = number of measurements; `mis` = number of missing values; `m` = mean; `md` = median; `sd` = standard deviation; `mad` = median average deviation; `min` = minimum; `max` = maximum; `trend` = weight of depended variable regressed on time (values ~ `mt`).

**Value**

A list containing a data frame of descriptive statistics (descriptives); the cse design (design); the number of cases (N)

**Author(s)**

Juergen Wilbert

**See Also**

[overlap](#), [plot.scdf](#)

**Examples**

```
## Descriptive statistics for a study of three single-cases
describe(Grosche2011)

## Descriptives of a three phase design
describe(exampleABC)

## Write descriptive statistics to .csv-file
study <- describe(Waddell2011)
write.csv(study$descriptives, file = tempfile())
```

---

design

*Generate a single-case design matrix*

---

**Description**

Generates a parameter list used for generating multiple random single-cases. This is used within the `random_scdf` function and the `power_test` function and for other Monte-Carlo tasks.

**Usage**

```
design(
  n = 1,
  phase_design = list(A = 5, B = 15),
  trend = 0,
  level = list(0),
  slope = list(0),
  start_value = 50,
  s = 10,
  rtt = 0.8,
  extreme_prop = list(0),
  extreme_range = c(-4, -3),
  missing_prop = 0,
  distribution = c("normal", "gaussian", "poisson", "binomial"),
```

```

n_trials = NULL,
mt = NULL,
B_start = NULL,
m,
phase.design,
MT,
B.start,
extreme.p,
extreme.d,
missing.p
)

```

### Arguments

- n** Number of cases to be designed (Default is  $n = 1$ ).
- phase\_design, phase.design** A list defining the length and label of each phase. E.g., `phase.length = list(A1 = 10, B1 = 10, A2 = 10, B2 = 10)`. Use vectors if you want to define different values for each case `phase.length = list(A = c(10, 15), B = c(10, 15))`.
- trend** Defines the effect size of a trend added incrementally to each measurement across the whole data-set. To assign different trends to several single-cases, use a vector of values (e.g. `trend = c(.1, .3, .5)`). If the number of cases exceeds the length of the vector, values are recycled. When using a 'gaussian' distribution, the trend parameters indicate effect size  $d$  changes. When using a binomial or poisson distribution, trend indicates an increase in points / counts per measurement.
- level** A list that defines the level increase (effect size  $d$ ) at the beginning of each phase relative to the previous phase (e.g. `list(A = 0, B = 1)`). The first element must be zero as the first phase of a single-case has no level effect (if you have one less list element than the number of phases, scan will add a leading element with 0 values). Use vectors to define variable level effects for each case (e.g. `list(A = c(0, 0), B = c(1, 2))`). When using a 'gaussian' distribution, the level parameters indicate effect size  $d$  changes. When using a binomial or poisson distribution, level indicates an increase in points / counts with the onset of each phase.
- slope** A list that defines the increase per measurement for each phase compared to the previous phase. `slope = list(A = 0, B = .1)` generates an incremental increase of 0.1 per measurement starting at the B phase. The first list element must be zero as the first phase of a single-case has no slope effect (if you have one less list element than the number of phases, scan will add a leading element with 0 values). Use vectors to define variable slope effects for each case (e.g. `list(A = c(0, 0), B = c(0.1, 0.2))`). If the number of cases exceeds the length of the vector, values are recycled. When using a 'gaussian' distribution, the slope parameters indicate effect size  $d$  changes per measurement. When using a binomial or poisson distribution, slope indicates an increase in points / counts per measurement.
- start\_value, m** Starting value at the first measurement. Default is 50. When `distribution = "poission"` the `start_value` represents frequency. When `distribution = "binomial"`

	start_value must range between 0 and 1 and they represent the probability of on event. To assign different start values to several single-cases, use a vector of values (e.g. <code>c(50, 42, 56)</code> ). If the number of cases exceeds the length of the vector, values are recycled. The 'm' argument is deprecated.
s	Standard deviation used to calculate absolute values from level, slope, trend effects and to calculate and error distribution from the rtt values. Set to 10 by default. To assign different variances to several single-cases, use a vector of values (e.g. <code>s = c(5, 10, 15)</code> ). If the number of cases exceeds the length of the vector, values are recycled. if the distribution is 'poisson' or 'binomial' s is not applied.
rtt	Reliability of the underlying simulated measurements. Set <code>rtt = .8</code> by default. To assign different reliabilities to several single-cases, use a vector of values (e.g. <code>rtt = c(.6, .7, .8)</code> ). If the number of cases exceeds the length of the vector, values are repeated. rtt has no effect when you're using binomial or poisson distributions.
extreme_prop, extreme.p	Probability of extreme values. <code>extreme.p = .05</code> gives a five percent probability of an extreme value. A vector of values assigns different probabilities to multiple cases. If the number of cases exceeds the length of the vector, values are recycled.
extreme_range, extreme.d	Range for extreme values. <code>extreme_range = c(-7, -6)</code> uses extreme values within a range of -7 and -6 . In case of a binomial or poisson distribution, <code>extreme_range</code> indicates frequencies. In case of a gaussian (or normal) distribution it indicates effect size d. Caution: the first value must be smaller than the second, otherwise the procedure will fail.
missing_prop, missing.p	Portion of missing values. <code>missing_prop = 0.1</code> creates 10% of all values as missing). A vector of values assigns different probabilities to multiple cases. If the number of cases exceeds the length of the vector, values are repeated.
distribution	Distribution of the criteria variable. Default is "normal". Possible values are "normal", "binomial", and "poisson".
n_trials	If distribution (see below) is "binomial", <code>n_trials</code> is the number of trials/observations/items.
mt, MT	Number of measurements (in each study). Default is <code>mt = 20</code> .
B_start, B.start	Phase B starting point. The default setting <code>B_start = 6</code> would assign the first five scores (of each case) to phase A, and all following scores to phase B. To assign different starting points for a set of multiple single-cases, use a vector of starting values (e.g. <code>B_start = c(6, 7, 8)</code> ). If the number of cases exceeds the length of the vector, values will be recycled.

## Value

An object of class `sc_design`.

**Author(s)**

Juergen Wibert

**Examples**

```
## Create random single-case data and inspect it
design <- design(
  n = 3, rtt = 0.75, slope = 0.1, extreme_prop = 0.1,
  missing_prop = 0.1
)
dat <- random_scdf(design, round = 1, random.names = TRUE, seed = 123)
describe(dat)

## And now have a look at poisson-distributed data
design <- design(
  n = 3, B_start = c(6, 10, 14), mt = c(12, 20, 22), start_value = 10,
  distribution = "poisson", level = -5, missing_prop = 0.1
)
dat <- random_scdf(design, seed = 1234)
pand(dat, decreasing = TRUE, correction = FALSE)
```

---

`estimate_design`*Estimate single-case design*

---

**Description**

This functions takes an scdf and extracts design parameters. The resulting object can be used to randomly create new scdf files with the same underlying parameters. This is useful for Monte-Carlo studies and bootstrapping procedures.

**Usage**

```
estimate_design(
  data,
  dvar,
  pvar,
  mvar,
  s = NULL,
  rtt = NULL,
  overall_effects = FALSE,
  overall_rtt = TRUE,
  model = "JW",
  ...
)
```

**Arguments**

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
<code>mvar</code>	Character string with the name of the measurement time variable. Defaults to the attributes in the scdf file.
<code>s</code>	The standard deviation depicting the between case variance of the overall performance. If more than two single-cases are included in the scdf, the variance is estimated if <code>s</code> is set to <code>NULL</code> .
<code>rtt</code>	The reliability of the measurements. The reliability is estimated when <code>rtt = NULL</code> .
<code>overall_effects</code>	If <code>TRUE</code> , trend, level, and slope effect estimations will be identical for each case. If <code>FALSE</code> , effects are estimated for each case separately.
<code>overall_rtt</code>	Ignored when 'rtt' is set. If <code>TRUE</code> , rtt estimations will be based on all cases and identical for each case. If <code>FALSE</code> rtt is estimated for each case separately.
<code>model</code>	Model used for calculating the dummy parameters (see Huitema & McKean, 2000). Default is <code>model = "W"</code> . Possible values are: "B&L-B", "H-M", "W", and deprecated "JW".
<code>...</code>	Further arguments passed to the <code>plm</code> function used for parameter estimation.

**Value**

A list of parameters for each single-case. Parameters include name, length, and starting measurement time of each phase, trend, level, and slope effects for each phase, start value, standard deviation, and reliability for each case.

**Examples**

```
# create a random scdf with predefined parameters
set.seed(1234)
design <- design(
  n = 10, trend = -0.02,
  level = list(0, 1), rtt = 0.8,
  s = 1
)
scdf<- random_scdf(design)

# Estimate the parameters based on the scdf and create a new random scdf
# based on these estimations
design_est <- estimate_design(scdf, rtt = 0.8)
scdf_est <- random_scdf(design_est)

# Analyze both datasets with an hplm model. See how similar the estimations
# are:
```

```

hplm(scdf, slope = FALSE)
hplm(scdf_est, slope = FALSE)

# Also similar results for pand and randomization tests:
pand(scdf)
pand(scdf_est)
rand_test(scdf)
rand_test(scdf_est)

```

---

export

*Export scan objects to html or latex*


---

### Description

This function is in an experimental status. Export creates html files of tables or displays them directly in the viewer pane of rstudio. When applied in rmarkdown/quarto, tables can also be created for pdf/latex output.

### Usage

```

export(object, ...)

## S3 method for class 'sc_desc'
export(
  object,
  caption = NA,
  footnote = NA,
  filename = NA,
  kable_styling_options = list(),
  kable_options = list(),
  flip = FALSE,
  ...
)

## S3 method for class 'sc_hplm'
export(
  object,
  caption = NA,
  footnote = NA,
  filename = NA,
  kable_styling_options = list(),
  kable_options = list(),
  round = 2,
  nice = TRUE,
  ...
)

## S3 method for class 'sc_nap'

```

```
export(  
  object,  
  caption = NA,  
  footnote = NA,  
  filename = NA,  
  kable_styling_options = list(),  
  kable_options = list(),  
  round = 2,  
  ...  
)  
  
## S3 method for class 'sc_overlap'  
export(  
  object,  
  caption = NA,  
  footnote = NA,  
  filename = NA,  
  kable_styling_options = list(),  
  kable_options = list(),  
  round = 2,  
  flip = FALSE,  
  ...  
)  
  
## S3 method for class 'sc_pem'  
export(  
  object,  
  caption = NA,  
  footnote = NA,  
  filename = NA,  
  kable_styling_options = list(),  
  kable_options = list(),  
  round = 2,  
  ...  
)  
  
## S3 method for class 'sc_plm'  
export(  
  object,  
  caption = NA,  
  footnote = NA,  
  filename = NA,  
  kable_styling_options = list(),  
  kable_options = list(),  
  round = 2,  
  nice = TRUE,  
  ...  
)
```

```
## S3 method for class 'sc_power'
export(
  object,
  caption = NA,
  footnote = NA,
  filename = NA,
  kable_styling_options = list(),
  kable_options = list(),
  ...
)

## S3 method for class 'sc_smd'
export(
  object,
  caption = NA,
  footnote = NA,
  filename = NA,
  select = c("Case", `Mean A` = "mA", `Mean B` = "mB", `SD A` = "sdA", `SD B` = "sdB",
    `SD Cohen` = "sd cohen", `SD Hedges` = "sd hedges", "Glass' delta", "Hedges' g",
    "Hedges' g correction", "Hedges' g durlak correction", "Cohen's d"),
  kable_styling_options = list(),
  kable_options = list(),
  round = 2,
  flip = FALSE,
  ...
)

## S3 method for class 'sc_tauu'
export(
  object,
  caption = NA,
  footnote = NA,
  filename = NA,
  select = "auto",
  kable_styling_options = list(),
  kable_options = list(),
  meta = TRUE,
  ...
)

## S3 method for class 'sc_trend'
export(
  object,
  caption = NA,
  footnote = NA,
  filename = NA,
  kable_styling_options = list(),
```

```

    kable_options = list(),
    round = 2,
    ...
  )

## S3 method for class 'scdf'
export(
  object,
  caption = NA,
  footnote = NA,
  filename = NA,
  kable_styling_options = list(),
  kable_options = list(),
  cols,
  ...
)

```

### Arguments

object	An scdf or an object exported from a scan function.
...	Further Arguments passed to internal functions.
caption	Character string with table caption. If left NA (default) a caption will be created based on the exported object.
footnote	Character string with table footnote. If left NA (default) a footnote will be created based on the exported object.
filename	String containing the file name. If a filename is given the output will be written to that file.
kable_styling_options	list with arguments passed to the kable_styling function.
kable_options	list with arguments passed to the kable function.
flip	If TRUE, some objects are exported with rows and columns flipped.
round	Integer passed to the digits argument internally used to round values.
nice	If set TRUE (default) output values are rounded and optimized for publication tables.
select	Character vector with name of variables to be included. When the vector is named, variables are renamed appropriately.
meta	If TRUE, the results of the meta analysis will be exported. If FALSE, each single-case is exported.
cols	Defines which columns are included when exporting an scdf. It is either a vector of variable names or the string "main" will select the central variables.

### Value

Returns or displays a specially formatted html (or latex) file.

---

fill_missing	<i>Replacing missing measurement times in single-case data</i>
--------------	----------------------------------------------------------------

---

### Description

The `fillmissingSC` function replaces missing measurements in single-case data.

### Usage

```
fill_missing(data, dvar, mvar, na.rm = TRUE)
```

### Arguments

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the <code>scdf</code> file.
<code>mvar</code>	Character string with the name of the measurement time variable. Defaults to the attributes in the <code>scdf</code> file.
<code>na.rm</code>	If set <code>TRUE</code> , NA values are also interpolated. Default is <code>na.rm = TRUE</code> .

### Details

This procedure is recommended if there are gaps between measurement times (e.g. MT: 1, 2, 3, 4, 5, ... 8, 9) or explicitly missing values in your single-case data and you want to calculate overlap indices ([overlapSC](#)) or a randomization test ([randSC](#)).

### Value

A single-case data frame with interpolated missing data points. See [scdf](#) to learn about the SCDF Format.

### Author(s)

Juergen Wilbert

### See Also

Other data manipulation functions: [add\\_l2\(\)](#), [as.data.frame.scdf\(\)](#), [as\\_scdf\(\)](#), [moving\\_median\(\)](#), [outlier\(\)](#), [ranks\(\)](#), [scdf](#), [select\\_cases\(\)](#), [set\\_vars\(\)](#), [shift\(\)](#), [smooth\\_cases\(\)](#), [standardize\(\)](#), [truncate\\_phase\(\)](#)

**Examples**

```
## In his study, Grosche (2011) could not realize measurements each single week for
## all participants. During the course of 100 weeks, about 20 measurements per person
## at different times were administered.

## Fill missing values in a single-case dataset with discontinuous measurement times
Grosche2011filled <- fill_missing(Grosche2011)
study <- c(Grosche2011[2], Grosche2011filled[2])
names(study) <- c("Original", "Filled")
plot(study)

## Fill missing values in a single-case dataset that are NA
Maggie <- random_scdf(design(level = list(0,1)), seed = 123)
Maggie_n <- Maggie
replace.positions <- c(10,16,18)
Maggie_n[[1]][replace.positions,"values"] <- NA
Maggie_f <- fill_missing(Maggie_n)
study <- c(Maggie, Maggie_n, Maggie_f)
names(study) <- c("original", "missing", "interpolated")
plot(study, marks = list(positions = replace.positions), style = "grid2")
```

---

hplm

*Hierarchical piecewise linear model / piecewise regression*


---

**Description**

The `hplm` function computes a hierarchical piecewise regression model.

**Usage**

```
hplm(
  data,
  dvar,
  pvar,
  mvar,
  model = c("W", "H-M", "B&L-B", "JW"),
  contrast = c("first", "preceding"),
  contrast_level = NA,
  contrast_slope = NA,
  method = c("ML", "REML"),
  control = list(opt = "optim"),
  random.slopes = FALSE,
  lr.test = FALSE,
  ICC = TRUE,
  trend = TRUE,
  level = TRUE,
```

```

    slope = TRUE,
    fixed = NULL,
    random = NULL,
    update.fixed = NULL,
    data.l2 = NULL,
    ...
)

```

## Arguments

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the <code>scdf</code> file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the <code>scdf</code> file.
<code>mvar</code>	Character string with the name of the measurement time variable. Defaults to the attributes in the <code>scdf</code> file.
<code>model</code>	Model used for calculating the dummy parameters (see Huitema & McKean, 2000). Default is <code>model = "W"</code> . Possible values are: "B&L-B", "H-M", "W", and deprecated "JW".
<code>contrast</code>	Sets <code>contrast_level</code> and <code>contrast_slope</code> . Either "first", "preceding" or a contrast matrix.
<code>contrast_level</code>	Either "first", "preceding" or a contrast matrix. If NA <code>contrast_level</code> is a copy of <code>contrast</code> .
<code>contrast_slope</code>	Either "first", "preceding" or a contrast matrix. If NA <code>contrast_level</code> is a copy of <code>contrast</code> .
<code>method</code>	Method used to fit your model. Pass "REML" to maximize the restricted log-likelihood or "ML" for maximized log-likelihood. Default is "ML".
<code>control</code>	A list of settings for the estimation algorithm, replacing the default values passed to the function <code>lmeControl</code> of the <code>nlme</code> package.
<code>random.slopes</code>	If <code>random.slopes = TRUE</code> random slope effects of the level, trend, and treatment parameter are estimated.
<code>lr.test</code>	If set <code>TRUE</code> likelihood ratio tests are calculated comparing model with vs. without random slope parameters.
<code>ICC</code>	If <code>ICC = TRUE</code> an intraclass-correlation is estimated.
<code>trend</code>	A logical indicating if a trend parameters is included in the model.
<code>level</code>	A logical indicating if a level parameters is included in the model.
<code>slope</code>	A logical indicating if a slope parameters is included in the model.
<code>fixed</code>	Defaults to the fixed part of the standard piecewise regression model. The parameter phase followed by the phase name (e.g., <code>phaseB</code> ) indicates the level effect of the corresponding phase. The parameter 'inter' followed by the phase name (e.g., <code>interB</code> ) addresses the slope effect based on the method provide in the model argument (e.g., "B&L-B"). The formula can be changed for example to include further L1 or L2 variables into the regression model.

random	The random part of the model.
update.fixed	An easier way to change the fixed model part (e.g., <code>. ~ . + newvariable</code> ).
data.l2	A dataframe providing additional variables at Level 2. The scdf File has to have names for all cases and the Level 2 dataframe has to have a column named 'cases' with the names of the cases the Level 2 variables belong to.
...	Further arguments passed to the lme function.

**Value**

model	List containing information about the applied model
N	Number of single-cases.
formula	A list containing the fixed and the random formulas of the hplm model.
hplm	Object of class lme containing the multilevel model
model.0	Object of class lme containing the Zero Model.
ICC	List containing intraclass correlation and test parameters.
model.without	Object of class gls containing the fixed effect model.

**Author(s)**

Juergen Wilbert

**See Also**

Other regression functions: [autocorr\(\)](#), [corrected\\_tau\(\)](#), [mplm\(\)](#), [plm\(\)](#), [trend\(\)](#)

**Examples**

```
## Compute hplm model on a MBD over fifty cases (restricted log-likelihood)
hplm(exampleAB_50, method = "REML", random.slopes = FALSE)

## Analyzing with additional L2 variables
Leidig2018 %>%
  add_l2(Leidig2018_l2) %>%
  hplm(update.fixed = .~. + gender + migration + ITRF_TOTAL*phaseB,
        slope = FALSE, random.slopes = FALSE, lr.test = FALSE
  )
```

---

is.scdf	<i>scdf objects Tests for objects of type "scdf"</i>
---------	------------------------------------------------------

---

**Description**

scdf objects Tests for objects of type "scdf"

**Usage**

```
is.scdf(x)
```

**Arguments**

x                    An object to be tested

**Value**

Returns TRUE or FALSE depending on whether its argument is of scdf type or not.

---

moving_median	<i>Transform every single case of a single case data frame</i>
---------------	----------------------------------------------------------------

---

**Description**

Takes an scdf and applies transformations to each individual case. This is useful to calculate or modify new variables.

**Usage**

```
moving_median(x, lag = 1)

moving_mean(x, lag = 1)

local_regression(x, mt = 1:length(x), f = 0.2)

first_of(x, positions = 0)

across_cases(...)

all_cases(...)

## S3 method for class 'scdf'
transform(`_data`, ...)
```

**Arguments**

x	A logical vector.
lag	Number of values surrounding a value to calculate the average
mt	A vector with measurement times.
f	the proportion of surrounding data influencing each data point.
positions	A numeric vector with relative positions to the first appearance of a TRUE value in x.
...	Expressions.
_data	An scdf.

**Details**

This function is a method of the generic transform function. Unlike the generic function, it calculates expressions serially. This means that the results of the calculation of one expression are the basis for the following computations. The `n` function returns the number of measurements in a case. The `all_cases` function is a helper function that extracts the values of a variable from all cases. It takes an expression as an argument. For example, `mean(all_cases(values))` calculates the mean of the values from all cases. `mean(all_cases(values[phase == "A"]))` will calculate the mean of all values where phase is A. The function `across_cases` allows to calculate new variables or replace existing variables across all cases. E.g., `across_cases(values_ranked = rank(values, na.last = "keep"))` will calculate a new variable with values ranked across all cases.

**Value**

An scdf.

**See Also**

Other data manipulation functions: [add\\_l2\(\)](#), [as.data.frame.scdf\(\)](#), [as\\_scdf\(\)](#), [fill\\_missing\(\)](#), [outlier\(\)](#), [ranks\(\)](#), [scdf](#), [select\\_cases\(\)](#), [set\\_vars\(\)](#), [shift\(\)](#), [smooth\\_cases\(\)](#), [standardize\(\)](#), [truncate\\_phase\(\)](#)

**Examples**

```
## Creates a single-case with frequency distributions. The proportion and
## percentage of the frequencies are calculated with transform:
design <- design(
  n = 3,
  level = 5,
  distribution = "binomial",
  n_trials = 20,
  start_value = 0.5
)
study <- random_scdf(design)
transform(study, proportion = values/trials, percentage = proportion * 100)

## Z standardize the dependent variable and add two new variables:
exampleAB %>%
```

```

transform(
  values = scale(values),
  mean_values = mean(values),
  sd_values = sd(values)
)

## Use `all` to calculate global variables.
exampleAB %>%
  transform(
    values_center_case = values - mean(values[phase == "A"]),
    values_center_global = values - mean(all(values[phase == "A"])),
    value_dif = values_center_case - values_center_global
  )

## Use `across_cases` to calculate or replace a variable with values from
## all cases. E.g., standardize the dependent variable:
exampleABC %>%
  transform(
    across_cases(values = scale(values))
  )

## Rank transform the values based on all cases vs. within each case:
exampleABC %>%
  transform(
    across_cases(values_across = rank(values, na.last="keep")),
    value_within = rank(values, na.last="keep")
  )

## Three helper functions to smooth the data
Huber2014$Berta %>%
  transform(
    "compliance (moving median)" = moving_median(compliance),
    "compliance (moving mean)" = moving_mean(compliance),
    "compliance (local regression)" = local_regression(compliance, mt)
  )

## Function first_of() helps to set NAs for specific phases.
## E.g., you want to replace the first two values of phase A and the first
## value of phase B and its preceding value.

byHeart2011 %>%
  transform(
    values = replace(values, first_of(phase == "A", 0:1), NA),
    values = replace(values, first_of(phase == "B", -1:0), NA)
  )

```

## Description

The `mplm` function computes a multivariate piecewise regression model.

**Usage**

```

mplm(
  data,
  dvar,
  mvar,
  pvar,
  model = c("W", "H-M", "B&L-B", "JW"),
  contrast = c("first", "preceding"),
  contrast_level = c(NA, "first", "preceding"),
  contrast_slope = c(NA, "first", "preceding"),
  trend = TRUE,
  level = TRUE,
  slope = TRUE,
  formula = NULL,
  update = NULL,
  na.action = na.omit,
  ...
)

```

**Arguments**

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
<code>mvar</code>	Character string with the name of the measurement time variable. Defaults to the attributes in the scdf file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
<code>model</code>	Model used for calculating the dummy parameters (see Huitema & McKean, 2000). Default is <code>model = "W"</code> . Possible values are: "B&L-B", "H-M", "W", and deprecated "JW".
<code>contrast</code>	Sets <code>contrast_level</code> and <code>contrast_slope</code> . Either "first", "preceding" or a contrast matrix.
<code>contrast_level</code>	Either "first", "preceding" or a contrast matrix. If NA <code>contrast_level</code> is a copy of <code>contrast</code> .
<code>contrast_slope</code>	Either "first", "preceding" or a contrast matrix. If NA <code>contrast_level</code> is a copy of <code>contrast</code> .
<code>trend</code>	A logical indicating if a trend parameters is included in the model.
<code>level</code>	A logical indicating if a level parameters is included in the model.
<code>slope</code>	A logical indicating if a slope parameters is included in the model.
<code>formula</code>	Defaults to the standard piecewise regression model. The parameter phase followed by the phase name (e.g., <code>phaseB</code> ) indicates the level effect of the corresponding phase. The parameter 'inter' followed by the phase name (e.g., <code>interB</code> ) addresses the slope effect based on the method provide in the model argument (e.g., "B&L-B"). The formula can be changed for example to include further variables into the regression model.

update	An easier way to change the regression formula (e.g., <code>. ~ . + newvariable</code> ).
na.action	Defines how to deal with missing values
...	Further arguments passed to the <code>lm</code> function.

**Value**

model	Character string from function call (see Arguments above).
full.model	Full regression model list

**Author(s)**

Juergen Wilbert

**See Also**

Other regression functions: [autocorr\(\)](#), [corrected\\_tau\(\)](#), [hplm\(\)](#), [plm\(\)](#), [trend\(\)](#)

**Examples**

```
res <- mplm(Leidig2018$`1a1`, dvar = c("academic_engagement", "disruptive_behavior"))
print(res)
## also report standardized coefficients:
print(res, std = TRUE)
```

---

 nap

---

*Nonoverlap of all Pairs*


---

**Description**

The `nap` function calculates the nonoverlap of all pairs (NAP; Parker & Vannest, 2009). NAP summarizes the overlap between all pairs of phase A and phase B data points. If an increase of phase B scores is expected, a non-overlapping pair has a higher phase B data point. The NAP equals  $\text{number of pairs showing no overlap} / \text{number of pairs}$ . Because NAP can only take values between 50 and 100 percent, a rescaled and therefore more intuitive NAP (0-100%) is also displayed.

**Usage**

```
nap(data, dvar, pvar, decreasing = FALSE, phases = c(1, 2))
```

**Arguments**

data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
dvar	Character string with the name of the dependent variable. Defaults to the attributes in the <code>scdf</code> file.
pvar	Character string with the name of the phase variable. Defaults to the attributes in the <code>scdf</code> file.

decreasing	If you expect data to be lower in the B phase, set decreasing = TRUE. Default is decreasing = FALSE.
phases	A vector of two characters or numbers indicating the two phases that should be compared. E.g., phases = c("A", "C") or phases = c(2, 4) for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., phases = list(A = c(1, 3), B = c(2, 4)) will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is phases = c(1, 2).

**Value**

nap	A data frame with NAP and additional values for each case.
N	Number of cases.

**Author(s)**

Juergen Wilbert

**References**

Parker, R. I., & Vannest, K. (2009). An improved effect size for single-case research: Nonoverlap of all pairs. *Behavior Therapy, 40*, 357-367.

**See Also**

Other overlap functions: [corrected\\_tau\(\)](#), [overlap\(\)](#), [pand\(\)](#), [pem\(\)](#), [pet\(\)](#), [pnd\(\)](#), [tau\\_u\(\)](#)

**Examples**

```
## Calculate NAP for a study with lower expected phase B scores
## (e.g. aggressive behavior)
gretchen <- scdf(c(A = 12, 14, 9, 10, B = 10, 6, 4, 5, 3, 4))
nap(gretchen, decreasing = TRUE)

## Request NAP for all cases from the Grosche2011 scdf
nap(Grosche2011)
```

---

outlier

*Handling outliers in single-case data*

---

**Description**

Identifies and drops outliers within a single-case data frame (scdf).

**Usage**

```

outlier(
  data,
  dvar,
  pvar,
  mvar,
  method = c("MAD", "Cook", "SD", "CI"),
  criteria = 3.5
)

```

**Arguments**

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
<code>mvar</code>	Character string with the name of the measurement time variable. Defaults to the attributes in the scdf file.
<code>method</code>	Specifies the method for outlier identification. Set <code>method = "MAD"</code> for mean average deviation, <code>method = "SD"</code> for standard deviations, <code>method = "CI"</code> for confidence intervals, <code>method = "Cook"</code> for Cook's Distance based on the Piecewise Linear Regression Model.
<code>criteria</code>	Specifies the criteria for outlier identification. Based on the method setting.

**Details**

For `method = "SD"`, `criteria = 2` would refer to two standard deviations. For `method = "MAD"`, `criteria = 3.5` would refer to 3.5 times the mean average deviation. For `method = "CI"`, `criteria = 0.99` would refer to a 99 percent confidence interval. For `method = "cook"`, `criteria = "4/n"` would refer to a Cook's Distance greater than  $4/n$ .

**Value**

<code>data</code>	A single-case data frame with substituted outliers.
<code>dropped.n</code>	A list with the number of dropped data points for each single-case.
<code>dropped.mt</code>	A list with the measurement-times of dropped data points for each single-case (values are based on the <code>mt</code> variable of each single-case data frame).
<code>sd.matrix</code>	A list with a matrix for each case with values for the upper and lower boundaries based on the standard deviation.
<code>ci.matrix</code>	A list with a matrix for each single-case with values for the upper and lower boundaries based on the confidence interval.
<code>cook</code>	A list of Cook's Distances for each measurement of each single-case.
<code>criteria</code>	Criteria used for outlier analysis.
<code>N</code>	Number of single-cases.
<code>case.names</code>	Case identifier.

**Author(s)**

Juergen Wilbert

**See Also**

Other data manipulation functions: [add\\_l2\(\)](#), [as.data.frame.scdf\(\)](#), [as\\_scdf\(\)](#), [fill\\_missing\(\)](#), [moving\\_median\(\)](#), [ranks\(\)](#), [scdf](#), [select\\_cases\(\)](#), [set\\_vars\(\)](#), [shift\(\)](#), [smooth\\_cases\(\)](#), [standardize\(\)](#), [truncate\\_phase\(\)](#)

**Examples**

```
## Identify outliers using 1.5 standard deviations as criterion
susanne <- random_scdf(level = 1.0)
res_outlier <- outlier(susanne, method = "SD", criteria = 1.5)
plot(susanne, marks = res_outlier)

## Identify outliers in the original data from Grosche (2011)
## using Cook's Distance greater than 4/n as criterion
res_outlier <- outlier(Grosche2011, method = "Cook", criteria = "4/n")
plot(Grosche2011, marks = res_outlier)
```

---

 overlap

*Overlap indices for single-case data*


---

**Description**

The overlap function provides the most common overlap indices for single-case data and some additional statistics.

**Usage**

```
overlap(data, dvar, pvar, mvar, decreasing = FALSE, phases = c(1, 2))
```

**Arguments**

data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
dvar	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
pvar	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
mvar	Character string with the name of the measurement time variable. Defaults to the attributes in the scdf file.
decreasing	If you expect data to be lower in the B phase, set decreasing = TRUE. Default is decreasing = FALSE.

phases A vector of two characters or numbers indicating the two phases that should be compared. E.g., `phases = c("A", "C")` or `phases = c(2, 4)` for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., `phases = list(A = c(1, 3), B = c(2, 4))` will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is `phases = c(1, 2)`.

### Details

See corresponding functions of PND, PEM, PET, NAP, PAND for calculation. `Tau_U` reports "A vs. B + Trend B - Trend A". `Base_Tau` is baseline corrected tau (correction applied when autocorrelation in phase A is significant). `Diff_mean` is the mean difference. `Diff_trend` is the difference in the regression estimation of the dependent variable on measurement-time ( $x \sim mt$ ) for each phase. `SMD` is the mean difference divided by the standard deviation of phase A. `Hedges_g` is the mean difference divided by the pooled standard deviation [ $\sqrt{((nA - 1) * sdA^2 + (nB - 1) * sdB^2) / (nA + nB - 2)}$ ] with a hedges correction applied [ $(1 - (3 / (4 * n - 9)))$ ].

### Value

`overlap` A data frame consisting of the following indices for each single-case for all cases: PND, PEM, PET, NAP, PAND, Tau-U (A vs. B - Trend A), `Diff_mean`, `Diff_trend`, SMD, Hedges-g.

`phases.A` Selection for A phase.

`phases.B` Selection for B phase.

`design` Phase design.

### Author(s)

Juergen Wilbert

### See Also

Other overlap functions: [corrected\\_tau\(\)](#), [nap\(\)](#), [pand\(\)](#), [pem\(\)](#), [pet\(\)](#), [pnd\(\)](#), [tau\\_u\(\)](#)

### Examples

```
## Display overlap indices for one single-case
overlap(Huitema2000, decreasing = TRUE)

## Display overlap indices for six single-cases
overlap(GruenkeWilbert2014)

## Combining phases for analyzing designs with more than two phases
overlap(exampleA1B1A2B2, phases = list(c("A1", "A2"), c("B1", "B2")))
```

---

pand *Percentage of all non-overlapping data*

---

### Description

The `pand` function calculates the percentage of all non-overlapping data (PAND; Parker, Hagan-Burke, & Vannest, 2007), an index to quantify a level increase (or decrease) in performance after the onset of an intervention.

### Usage

```
pand(data, dvar, pvar, decreasing = FALSE, correction = TRUE, phases = c(1, 2))
```

### Arguments

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the <code>scdf</code> file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the <code>scdf</code> file.
<code>decreasing</code>	If you expect data to be lower in the B phase, set <code>decreasing = TRUE</code> . Default is <code>decreasing = FALSE</code> .
<code>correction</code>	The default <code>correction = TRUE</code> makes <code>pand</code> use a frequency matrix, which is corrected for ties. A tie is counted as the half of a measurement in both phases. Set <code>correction = FALSE</code> to use the uncorrected matrix, which is not recommended.
<code>phases</code>	A vector of two characters or numbers indicating the two phases that should be compared. E.g., <code>phases = c("A", "C")</code> or <code>phases = c(2, 4)</code> for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., <code>phases = list(A = c(1, 3), B = c(2, 4))</code> will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is <code>phases = c(1, 2)</code> .

### Details

The PAND indicates nonoverlap between phase A and B data (like PND), but uses all data and is therefore not based on one single (probably unrepresentative) datapoint. Furthermore, PAND allows the comparison of real and expected associations (Chi-square test) and estimation of the effect size Phi, which equals Pearson's  $r$  for dichotomous data. Thus, phi-Square is the amount of explained variance. The original procedure for computing the PAND (Parker, Hagan-Burke, & Vannest, 2007) does not account for ambivalent datapoints (ties). The newer NAP overcomes this problem and has better precision-power (Parker, Vannest, & Davis, 2014).

**Value**

pand	Percentage of all non-overlapping data.
phi	Effect size Phi based on expected and observed values.
perc_overlap	Percentage of overlapping data points.
overlaps	Number of overlapping data points.
n	Number of data points.
N	Number of cases.
nA	Number of data points in phase A.
nB	Number of data points in phase B.
pA	Percentage of data points in phase A.
pB	Percentage of data points in phase B.
matrix	2x2 frequency matrix of phase A and B comparisons.
matrix_counts	2x2 counts matrix of phase A and B comparisons.
correlation	A list of the correlation values: statistic, parameter, p.value, estimate, null.value, alternative, method, data.name, correction.
correction	Logical argument from function call (see Arguments above).

**Author(s)**

Juergen Wilbert

**References**

- Parker, R. I., Hagan-Burke, S., & Vannest, K. (2007). Percentage of All Non-Overlapping Data (PAND): An Alternative to PND. *The Journal of Special Education, 40*, 194-204.
- Parker, R. I., & Vannest, K. (2009). An Improved Effect Size for Single-Case Research: Nonoverlap of All Pairs. *Behavior Therapy, 40*, 357-367.

**See Also**

Other overlap functions: [corrected\\_tau\(\)](#), [nap\(\)](#), [overlap\(\)](#), [pem\(\)](#), [pet\(\)](#), [pnd\(\)](#), [tau\\_u\(\)](#)

**Examples**

```
## Calculate the PAND for a MMBD over three cases
gunnar <- scdf(c(2,3,1,5,3,4,2,6,4,7), B_start = 5)
birgit <- scdf(c(3,3,2,4,7,4,2,1,4,7), B_start = 4)
bodo <- scdf(c(2,3,4,5,3,4,7,6,8,7), B_start = 6)
mbd <- c(gunnar, birgit, bodo)
pand(mbd)
pand(bodo)

## Calculate the PAND with an expected decrease of phase B scores
cubs <- scdf(c(20,22,24,17,21,13,10,9,20,9,18), B_start = 5)
pand(cubs, decreasing = TRUE)
```

pem

*Percent exceeding the median***Description**

The pem function returns the percentage of phase B data exceeding the phase A median. Additionally, a chi square test against a 50/50 distribution is computed. Different measures of central tendency can be addressed for alternative analyses.

**Usage**

```
pem(
  data,
  dvar,
  pvar,
  decreasing = FALSE,
  binom.test = TRUE,
  chi.test = FALSE,
  FUN = median,
  phases = c(1, 2),
  ...
)
```

**Arguments**

data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
dvar	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
pvar	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
decreasing	If you expect data to be lower in the B phase, set decreasing = TRUE. Default is decreasing = FALSE.
binom.test	Computes a binomial test for a 50/50 distribution. Default is binom.test = TRUE.
chi.test	Computes a Chi-square test. The default setting chi.test = FALSE skips the Chi-square test.
FUN	Data points are compared with the phase A median. Use this argument to implement alternative measures of central tendency. Default is FUN = median
phases	A vector of two characters or numbers indicating the two phases that should be compared. E.g., phases = c("A", "C") or phases = c(2, 4) for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., phases = list(A = c(1, 3), B = c(2, 4)) will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is phases = c(1, 2).

... Additional arguments for the FUN parameter (e.g. FUN = mean, trim = 0.1 will use the 10 percent trimmed arithmetic mean instead of the median for comparisons). The function must take a vector of numeric values and the na.rm argument and return a numeric value.

### Author(s)

Juergen Wilbert

### See Also

Other overlap functions: [corrected\\_tau\(\)](#), [nap\(\)](#), [overlap\(\)](#), [pand\(\)](#), [pet\(\)](#), [pnd\(\)](#), [tau\\_u\(\)](#)

### Examples

```
## Calculate the PEM including the Binomial and Chi-square tests for a single-case
dat <- random_scdf(5, level = 0.5)
pem(dat, chi.test = TRUE)
```

---

pet	<i>Percent exceeding the trend</i>
-----	------------------------------------

---

### Description

The pet function provides the percentage of phase B data points exceeding the prediction based on the phase A trend. A binomial test against a 50/50 distribution is computed. Furthermore, the percentage of phase B data points exceeding the upper (or lower) 95 percent confidence interval of the predicted progress is computed.

### Usage

```
pet(data, dvar, pvar, mvar, ci = 0.95, decreasing = FALSE, phases = c(1, 2))
```

### Arguments

data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
dvar	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
pvar	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
mvar	Character string with the name of the measurement time variable. Defaults to the attributes in the scdf file.
ci	Width of the confidence interval. Default is ci = 0.95.
decreasing	If you expect data to be lower in the B phase, set decreasing = TRUE. Default is decreasing = FALSE.

phases A vector of two characters or numbers indicating the two phases that should be compared. E.g., `phases = c("A", "C")` or `phases = c(2, 4)` for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., `phases = list(A = c(1, 3), B = c(2, 4))` will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is `phases = c(1, 2)`.

### Value

PET	Percent exceeding the trend.
PET.ci	Percent exceeding the upper / lower 95%-CI boundary.
p	P value of Binomial Test.
ci.percent	Width of confidence interval in percent.
se.factors	Standard error.
N	Number of cases.
decreasing	Logical argument from function call (see Arguments above).
case.names	Assigned name of single-case.
phases	-

### Author(s)

Juergen Wilbert

### See Also

Other overlap functions: [corrected\\_tau\(\)](#), [nap\(\)](#), [overlap\(\)](#), [pand\(\)](#), [pem\(\)](#), [pnd\(\)](#), [tau\\_u\(\)](#)

### Examples

```
## Calculate the PET and use a 99%-CI for the additional calculation
# create random example data
design <- design(n = 5, slope = 0.2)
dat <- random_scdf(design, seed = 23)
pet(dat, ci = .99)
```

### Description

The `plm` function computes a piecewise regression model (see Huitema & McKean, 2000).

**Usage**

```
plm(
  data,
  dvar,
  pvar,
  mvar,
  AR = 0,
  model = c("W", "H-M", "B&L-B", "JW"),
  family = "gaussian",
  trend = TRUE,
  level = TRUE,
  slope = TRUE,
  contrast = c("first", "preceding"),
  contrast_level = c(NA, "first", "preceding"),
  contrast_slope = c(NA, "first", "preceding"),
  formula = NULL,
  update = NULL,
  na.action = na.omit,
  r_squared = TRUE,
  var_trials = NULL,
  dvar_percentage = FALSE,
  ...
)
```

**Arguments**

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the <code>scdf</code> file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the <code>scdf</code> file.
<code>mvar</code>	Character string with the name of the measurement time variable. Defaults to the attributes in the <code>scdf</code> file.
<code>AR</code>	Maximal lag of autoregression. Modeled based on the Autoregressive-Moving Average (ARMA) function. When <code>AR</code> is set, the <code>family</code> argument must be set to <code>family = "gaussian"</code> .
<code>model</code>	Model used for calculating the dummy parameters (see Huitema & McKean, 2000). Default is <code>model = "W"</code> . Possible values are: "B&L-B", "H-M", "W", and deprecated "JW".
<code>family</code>	Set the distribution family. Defaults to a gaussian distribution. See the <code>family</code> function for more details.
<code>trend</code>	A logical indicating if a trend parameters is included in the model.
<code>level</code>	A logical indicating if a level parameters is included in the model.
<code>slope</code>	A logical indicating if a slope parameters is included in the model.
<code>contrast</code>	Sets <code>contrast_level</code> and <code>contrast_slope</code> . Either "first", "preceding" or a contrast matrix.

contrast_level	Either "first", "preceding" or a contrast matrix. If NA contrast_level is a copy of contrast.
contrast_slope	Either "first", "preceding" or a contrast matrix. If NA contrast_level is a copy of contrast.
formula	Defaults to the standard piecewise regression model. The parameter phase followed by the phase name (e.g., phaseB) indicates the level effect of the corresponding phase. The parameter 'inter' followed by the phase name (e.g., interB) addresses the slope effect based on the method provide in the model argument (e.g., "B&L-B"). The formula can be changed for example to include further variables into the regression model.
update	An easier way to change the regression formula (e.g., . ~ . + newvariable).
na.action	Defines how to deal with missing values.
r_squared	Logical. If TRUE, delta r_squares will be calculated for each predictor.
var_trials	Name of the variable containing the number of trials (only for binomial regressions). If a single integer is provided this is considered to be a the constant number of trials across all measurements.
dvar_percentage	Only for binomial distribution. If set TRUE, the dependent variable is assumed to represent proportions [0,1]. Otherwise dvar is assumed to represent counts.
...	Further arguments passed to the glm function.

### Value

formula	plm formula. Useful if you want to use the update or formula argument and you don't know the names of the parameters.
model	Character string from function call (see Arguments above).
F.test	F-test values of modelfit.
r.squares	Explained variance R squared for each model parameter.
ar	Autoregression lag from function call (see Arguments above).
family	Distribution family from function call (see Arguments above).
full.model	Full regression model list from the gls or glm function.

### Author(s)

Juergen Wilbert

### References

- Beretvas, S., & Chung, H. (2008). An evaluation of modified R2-change effect size indices for single-subject experimental designs. *Evidence-Based Communication Assessment and Intervention*, 2, 120-128.
- Huitema, B. E., & McKean, J. W. (2000). Design specification issues in time-series intervention models. *Educational and Psychological Measurement*, 60, 38-58.

**See Also**

Other regression functions: [autocorr\(\)](#), [corrected\\_tau\(\)](#), [hplm\(\)](#), [mplm\(\)](#), [trend\(\)](#)

**Examples**

```
## Compute a piecewise regression model for a random single-case
set.seed(123)
AB <- design(
  phase_design = list(A = 10, B = 20),
  level = list(A = 0, B = 1), slope = list(A = 0, B = 0.05),
  trend = 0.05
)
dat <- random_scdf(design = AB)
plm(dat, AR = 3)

## Another example with a more complex design
A1B1A2B2 <- design(
  phase_design = list(A1 = 15, B1 = 20, A2 = 15, B2 = 20),
  level = list(A1 = 0, B1 = 1, A2 = -1, B2 = 1),
  slope = list(A1 = 0, B1 = 0.0, A2 = 0, B2 = 0.0),
  trend = 0.0)
dat <- random_scdf(design = A1B1A2B2, seed = 123)
plm(dat, contrast = "preceding")

## no slope effects were found. Therefore, you might want to the drop slope
## estimation:
plm(dat, slope = FALSE, contrast = "preceding")

## and now drop the trend estimation as well
plm(dat, slope = FALSE, trend = FALSE, contrast = "preceding")

## A poisson regression
example_A24 %>%
  plm(family = "poisson")

## A binomial regression (frequencies as dependent variable)
plm(exampleAB_score$Christiano, family = "binomial", var_trials = "trials")

## A binomial regression (percentage as dependent variable)
exampleAB_score$Christiano %>%
  transform(percentage = values/trials) %>%
  set_dvar("percentage") %>%
  plm(family = "binomial", var_trials = "trials", dvar_percentage = TRUE)
```

---

plot.scdf

*Plot single-case data*

---

**Description**

This function provides a plot of a single-case or multiple single-cases.

**Usage**

```
## S3 method for class 'scdf'
plot(...)

plotSC(
  data,
  dvar,
  pvar,
  mvar,
  ylim = NULL,
  xlim = NULL,
  xinc = 1,
  lines = NULL,
  marks = NULL,
  phase.names = NULL,
  xlab = NULL,
  ylab = NULL,
  main = "",
  case.names = NULL,
  style = getOption("scan.plot.style"),
  ...
)
```

**Arguments**

...	Further arguments passed to the plot command.
data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
dvar	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
pvar	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
mvar	Character string with the name of the measurement time variable. Defaults to the attributes in the scdf file.
ylim	Lower and upper limits of the y-axis (e.g., <code>ylim = c(0, 20)</code> sets the y-axis to a scale from 0 to 20). With multiple single-cases you can use <code>ylim = c(0, NA)</code> to scale the y-axis from 0 to the maximum of each case. <code>ylim</code> is not set by default, which makes scan set a proper scale based on the given data.
xlim	Lower and upper limits of the x-axis (e.g., <code>xlim = c(0, 20)</code> sets the x-axis to a scale from 0 to 20). With multiple single-cases you can use <code>ylim = c(0, NA)</code> to scale the x-axis from 0 to the maximum of each case. <code>xlim</code> is not set by default, which makes scan set a proper scale based on the given data.
xinc	An integer. Increment of the x-axis. 1 :each mt value will be printed, 2 : every other value, 3 : every third values etc.
lines	A list defining one or multiple lines or curves to be plotted. The argument is passed as a list (e.g., <code>list(type = "median")</code> ). Some of the procedures can be refined with an additional argument (e.g., <code>lines = list(type = "mean", trim</code>

= 0.2) adds a 20% trimmed mean line. For multiple lines, provide a list element for each line (e.g., `list(list(type = "median", col = "red"), list(type = "trend", col = "blue"))`). Possible lines are:

- "median" Separate lines for phase A and B medians.
- "mean" Separate lines for phase A and B means. By default it is 10%-trimmed. Other trims can be set, using a trim parameter (e.g., `lines = list(type = "mean", trim = 0.2)` draws a 20%-trimmed mean line).
- "trend" Separate lines for phase A and B trends.
- "trendA" OLS trend line for phase A, extrapolated throughout phase B.
- "trendA\_bisplit" Split middle (bi-split) trend line for phase A, extrapolated throughout phase B.
- "trendA\_trisplit" Tukey tri-split trend line for phase A, extrapolated throughout phase B.
- "maxA/minA" Line at the level of the highest or lowest phase A score.
- "medianA" Line at the phase A median score.
- "meanA" Line at the phase A 10%-trimmed mean score. Apply a different trim, by using the additional argument (e.g., `lines = list(type = "meanA", trim = 0.2)`).
- "plm" Regression lines for piecewise linear regression model.
- "plm.ar" Regression lines for piecewise autoregression model. The lag is specified like this: `lines = list(type = "plm.ar", ar = 2)`. Default lag is set to 2.
- "movingMean" Draws a moving mean curve, with a specified lag: `lines = list(type = "movingMean", lag = 2)`. Default is a lag 1 curve.
- "movingMedian" Draws a moving median curve, with a specified lag: `lines = list(type = "movingMedian", lag = 3)`. Default is a lag 1 curve.
- "loreg" Draws a non-parametric local regression line. The proportion of data influencing each data point can be specified using `lines = list(type = "loreg", mf = 0.66)`. The default is 0.5.
- "lty" Use this argument to define the line type. Examples are: "solid", "dashed", "dotted".
- "lwd" Use this argument to define the line's thickness, e.g., `lwd = 4`.
- "col" Use this argument to define the line's color, e.g., `col = "red"`.

marks

A list of parameters defining markings of certain data points.

- "positions" A vector or a list of vectors indicating measurement-times to be highlighted. In case of a vector, the marked measurement-times are the same for all plotted cases. In case of a list of vectors, marks are set differently for each case. The list must have the same length as there are cases in the data file.
- "col" Color of the marks.
- "cex" Size of the marks.

Use for example `marks = list(positions = c(1, 8, 15), col = "red", cex = 3)` to make the MTs one, eight and 18 appear big and red.

phase.names

By default phases are labeled based on the levels of the phase variable. Use this argument to specify different labels: `phase.names = c("Baseline", "Intervention")`.

xlab	The label of the x-axis. Default is xlab = "Measurement time".
ylab	The labels of the y-axis. Default is ylab = "Score".
main	Main title of the plot.
case.names	Case names. If not provided, names are taken from the scdf. Set case.names = "" if you don't like to include case names.
style	Either a character with the name of a pre-implemented style or a style object. See <a href="#">style_plot</a> to learn about this format.

**Value**

Returns a plot of one or multiple single-cases.

**Author(s)**

Juergen Wilbert

**See Also**

[style\\_plot](#), [describeSC](#), [overlapSC](#)

**Examples**

```
## Request the default plot of the data from Borckhardt (2014)
plot(Borckardt2014)

## Plot the three cases from Grosche (2011) and visualize the phase A trend
plot(Grosche2011, style = "grid", lines = "trendA")

## Request the local regression line for Georg from that data set and customize the plot
plot(Grosche2011$Georg, style = "sienna", ylim = c(0,NA),
     xlab = "Training session", ylab = "Words per minute",
     phase.names = c("Baseline", "Intervention"), xinc = 5,
     lines = list(type = "loreg", f = 0.2, lty = "solid", col = "black", lwd = 3))

## Plot a random MBD over three cases and mark interesting MTs
dat <- random_scdf(design = design(3))
plot(dat, marks = list(positions = list(c(2,4,5),c(1,2,3),c(7,8,9))), col = "blue",
     cex = 1.4), style = c("grid", "annotate", "tiny"))
```

---

plot\_rand

*Plot random distribution*

---

**Description**

This function takes the return of the `rand_test` function and creates a histogram with the distribution of the `rand` sample statistics.

**Usage**

```
plot_rand(
  object,
  xlab = NA,
  ylab = "Frequency",
  title = "Random distribution",
  text_observed = "observed",
  color = "lightgrey",
  ...
)
```

**Arguments**

object	Object returned from the <code>rand_test()</code> function
xlab	Label for the x-axis.
ylab	Label for the y-axis.
title	Plot title.
text_observed	Text for marking the number of observed statistic.
color	Bar color.
...	Further arguments passed to the plot function.

pnd

*Percentage of non-overlapping data***Description**

This function returns the percentage of non-overlapping data. Due to its error-proneness the PND should not be used, but [nap](#) or [pand](#) instead (see Parker & Vannest, 2009).

**Usage**

```
pnd(data, dvar, pvar, decreasing = FALSE, phases = c("A", "B"))
```

**Arguments**

data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
dvar	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
pvar	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
decreasing	If you expect data to be lower in the B phase, set <code>decreasing = TRUE</code> . Default is <code>decreasing = FALSE</code> .
phases	A vector of two characters or numbers indicating the two phases that should be compared. E.g., <code>phases = c("A", "C")</code> or <code>phases = c(2, 4)</code> for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., <code>phases = list(A = c(1, 3), B = c(2, 4))</code> will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is <code>phases = c(1, 2)</code> .

**Value**

PND                      Percentage of non-overlapping data.

**Author(s)**

Juergen Wilbert

**See Also**

Other overlap functions: [corrected\\_tau\(\)](#), [nap\(\)](#), [overlap\(\)](#), [pand\(\)](#), [pem\(\)](#), [pet\(\)](#), [tau\\_u\(\)](#)

**Examples**

```
## Calculate the PND for multiple single-case data
pnd(GruenkeWilbert2014)
```

---

power\_test

*Empirical power analysis for single-case data*

---

**Description**

Conducts a Monte-Carlo study on the test-power and alpha-error probability of a statistical function.

**Usage**

```
power_test(
  design,
  method = c("plm_level", "rand", "tauU"),
  effect = "level",
  n_sim = 100,
  design_is_one_study = TRUE,
  alpha_test = TRUE,
  power_test = TRUE,
  binom_test = FALSE,
  binom_test_alpha = FALSE,
  binom_test_power = FALSE,
  binom_test_correct = FALSE,
  ci = FALSE,
  alpha_level = 0.05
)
```

**Arguments**

design	An object returned from the 'design' function.
method	A (named) list that defines the methods the power analysis is based on. Each element can contain a function (that takes an scdf file and returns a p value) or a character string (the name of predefined functions). default method = list("plm_level", "rand", "tauU") computes a power analysis based on <a href="#">tau_u</a> , <a href="#">rand_test</a> and <a href="#">plm</a> analyses. (Further predefined functions are: "plm_slope", "plm_poisson_level", "plm_poisson_slope", "hplm_level", "hplm_slope", "base_tau").
effect	Either "level" or "slope". The respective effect of the provided design is set to 0 when computing the alpha-error proportion.
n_sim	Number of sample studies created for the the Monte-Carlo study. Default is n = 100. Ignored if design_is_one_study = FALSE.
design_is_one_study	If TRUE, the design is assumed to define all cases of one study that is repeatedly randomly created n_sim times. If false, the design is assumed to contain all cases from which a random sample is generated. This is useful for very specific complex simulation studies.
alpha_test	Logical. If TRUE, alpha error is calculated.
power_test	Logical. If TRUE, power is calculated.
binom_test	Shortcut. When set TRUE, binom_test_power is set to 0.80, binom_test_alpha is set to 0.05, and binom_test_correct is set to 0.875.
binom_test_alpha	Either FALSE or a value. If a value is provided, a binomial test is calculated testing if the alpha error proportion is less than the provided value.
binom_test_power	Either FALSE or a value. If a value is provided, a binomial test is calculated testing if the power is greater than the provided value.
binom_test_correct	Either FALSE or a value. If a value is provided, a binomial test is calculated testing if the correct proportion is greater than the provided value.
ci	Either FALSE or a value. If a value is provided, confidence intervals at the provided level are calculated for power, alpha error, and correct proportions.
alpha_level	Alpha level used to calculate the proportion of significant tests. Default is alpha_level = 0.05.

**Details**

Based on a [design](#) object, a large number of single-cases are generated and re-analyzed with a provided statistical function. The proportion of significant analyzes is the test power. In a second step, a specified effect of the design object is set to 0 and again single-cases are generated and reanalyzed. The proportion of significant analyzes is the alpha error probability.

**Author(s)**

Juergen Wilbert

**See Also**[random\\_scdf, design](#)**Examples**

```
## Assume you want to conduct a single-case study with 15 measurements
## (phases: A = 6 and B = 9) using a highly reliable test and
## an expected level effect of d = 1.4.
## A (strong) trend effect is trend = 0.05. What is the power?
## (Note: n_sims is set to 10. Set n_sims to 1000 for a serious calculation.)
design <- design(
  n = 1, phase_design = list(A = 6, B = 9),
  rtt = 0.8, level = 1.4, trend = 0.05
)
power_test(design, n_sim = 10)

## Would you achieve higher power by setting up a MBD with three cases?
design <- design(
  n = 3, phase_design = list(A = 6, B = 9),
  rtt = 0.8, level = 1.4, trend = 0.05
)
power_test(design, n_sim=10, method=list("hplm_level", "rand", "tauU_meta"))
```

print.scdf

*Print an scdf***Description**

Print an scdf

**Usage**

```
## S3 method for class 'scdf'
print(
  x,
  cases = getOption("scan.print.cases"),
  rows = getOption("scan.print.rows"),
  cols = getOption("scan.print.cols"),
  long = getOption("scan.print.long"),
  digits = getOption("scan.print.digits"),
  ...
)
```

**Arguments**

x                    An scdf object

cases                Number of cases to be printed. "fit" fits the number to the current screen width.

rows	Number of rows to be printed.
cols	Columns to be printed. "Main" only prints the dependent, measurement-time and phase variable.
long	Logical. If TRUE cases are printed in one by a time.
digits	Number of digits.
...	Further arguments passed to the print function.

### Details

Print options for scdf objects could be set globally: `option(scan.print.cases = "all")`, `option(scan.print.rows = 10)`, `option(scan.print.cols = "main")`, `option(scan.print.long = TRUE)`, `option(scan.print.digits = 0)`, `option(scan.print.scdf.name = FALSE)`

---

random_scdf	<i>Single-case data generator</i>
-------------	-----------------------------------

---

### Description

The `random_scdf` function generates random single-case data frames for monte-carlo studies and demonstration purposes. `design` is used to set up a design matrix with all parameters needed for the `random_scdf` function.

### Usage

```
random_scdf(design = NULL, round = NA, random_names = FALSE, seed = NULL, ...)
```

### Arguments

design	A design matrix which is created by <code>design</code> and specifies all parameters.
round	Rounds the scores to the defined decimal. To round to the second decimal, set <code>round = 2</code> .
random_names	Is FALSE by default. If set <code>random_names = TRUE</code> cases are assigned random first names. If set "neutral", "male" or "female" only gender neutral, male, or female names are chosen. The names are drawn from the 2,000 most popular names for newborns in 2012 in the U.S. (1,000 male and 1,000 female names).
seed	A seed number for the random generator.
...	arguments that are directly passed to the <code>design</code> function for a more concise coding.

### Value

A single-case data frame. See [scdf](#) to learn about this format.

### Author(s)

Juergen Wibert

**Examples**

```
## Create random single-case data and inspect it
design <- design(
  n = 3, rtt = 0.75, slope = 0.1, extreme_prop = 0.1,
  missing_prop = 0.1
)
dat <- random_scdf(design, round = 1, random_names = TRUE, seed = 123)
describe(dat)

## And now have a look at poisson-distributed data
design <- design(
  n = 3, B_start = c(6, 10, 14), mt = c(12, 20, 22), start_value = 10,
  distribution = "poisson", level = -5, missing_prop = 0.1
)
dat <- random_scdf(design, seed = 1234)
pand(dat, decreasing = TRUE, correction = FALSE)
```

---

rand\_test

*Randomization Tests for single-case data*


---

**Description**

The randSC function computes a randomization test for single or multiple baseline single-case data. The function is based on an algorithm from the SCRT package (Bulte & Onghena, 2009, 2012), but rewritten and extended for the use in AB designs.

**Usage**

```
rand_test(
  data,
  dvar,
  pvar,
  statistic = c("Mean B-A", "Mean A-B", "Median B-A", "Median A-B", "Mean |A-B|"),
  number = 500,
  complete = FALSE,
  limit = 5,
  startpoints = NA,
  exclude.equal = FALSE,
  phases = c(1, 2),
  graph = FALSE,
  output = NULL,
  seed = NULL
)
```

**Arguments**

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the <code>scdf</code> file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the <code>scdf</code> file.
<code>statistic</code>	<p>Defines the statistic on which the comparison of phases A and B is based on. Default setting is <code>statistic = "Mean B-A"</code>. The following comparisons are possible:</p> <ul style="list-style-type: none"> <li>• "Mean A-B": Uses the difference between the mean of phase A and the mean of phase B. This is appropriate if a decrease of scores was expected for phase B.</li> <li>• "Mean B-A": Uses the difference between the mean of phase B and the mean of phase A. This is appropriate if an increase of scores was expected for phase B.</li> <li>• "Mean  A-B ": Uses the absolute value of the difference between the means of phases A and B.</li> <li>• "Median A-B": The same as "Mean A-B", but based on the median.</li> <li>• "Median B-A": The same as "Mean B-A", but based on the median.</li> </ul>
<code>number</code>	Sample size of the randomization distribution. The exactness of the p-value can not exceed $1/\text{number}$ (i.e., <code>number = 100</code> results in p-values with an exactness of one percent). Default is <code>number = 500</code> . For faster processing use <code>number = 100</code> . For more precise p-values set <code>number = 1000</code> .
<code>complete</code>	If TRUE, the distribution is based on a complete permutation of all possible starting combinations. This setting overwrites the <code>number</code> Argument. The default setting is FALSE.
<code>limit</code>	Minimal number of data points per phase in the sample. The first number refers to the A-phase and the second to the B-phase (e.g., <code>limit = c(5,3)</code> ). If only one number is given, this number is applied to both phases. Default is <code>limit = 5</code> .
<code>startpoints</code>	Alternative to the <code>limit</code> -parameter <code>startpoints</code> exactly defines the possible start points of phase B (e.g., <code>startpoints = 4:9</code> restricts the phase B start points to measurements 4 to 9. <code>startpoints</code> overruns the <code>limit</code> -parameter.
<code>exclude.equal</code>	If set to <code>exclude.equal = FALSE</code> , which is the default, random distribution values equal to the observed distribution are counted as null-hypothesis conform. That is, they decrease the probability of rejecting the null-hypothesis (increase the p-value). <code>exclude.equal</code> should be set to TRUE if you analyse one single-case design (not a multiple baseline data set) to reach a sufficient power. But be aware, that it increases the chance of an alpha-error.
<code>phases</code>	A vector of two characters or numbers indicating the two phases that should be compared. E.g., <code>phases = c("A", "C")</code> or <code>phases = c(2,4)</code> for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., <code>phases = list(A = c(1,3), B = c(2,4))</code> will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is <code>phases = c(1,2)</code> .

graph	If graph = TRUE, a histogram of the resulting distribution is plotted. It is FALSE by default. <i>Note: use the more versatile <a href="#">plot_rand</a> function instead.</i>
output	(deprecated and not implemented)
seed	A seed number for the random generator.

**Value**

statistic	Character string from function call (see Arguments above).
N	Number of single-cases.
n1	Number of data points in phase A.
n2	Number of data points in phase B.
limit	Numeric from function call (see Arguments above).
startpoints	A vector defining the start points passed from the function call (see Arguments above).
p.value	P-value of the randomization test for the given data.
number	Sample size of randomization distribution from function call (see Arguments above).
complete	Logical argument from function call (see Arguments above).
observed.statistic	Test statistic observed for the given single-case data. (see statistic in the Arguments above.)
Z	Z-value of observed test statistic.
p.z.single	Probability of z-value.
distribution	Test statistic distribution from randomized data sets.
possible.combinations	Number of possible combinations under the given restrictions.
auto.corrected.number	TRUE indicates that a corrected number of combinations was used. This happens, if the number of possible combinations (under the given restrictions) undercuts the requested number of combinations.

**Author(s)**

Juergen Wilbert

**References**

- Bulte, I., & Onghena, P. (2009). Randomization tests for multiple-baseline designs: An extension of the SCRT-R package. *Behavior Research Methods*, *41*, 477-485.
- Bulte, I., & Onghena, P. (2012). *SCRT: Single-Case Randomization Tests*. Available from: <https://CRAN.R-project.org/package=SCRT>

## Examples

```
## Compute a randomization test on the first case of the byHeart2011 data and include a graph
rand_test(byHeart2011[1], statistic = "Median B-A", graph = TRUE, seed = 123)
```

```
## Compute a randomization test on the Grosche2011 data using complete permutation
rand_test(Grosche2011, statistic = "Median B-A", complete = TRUE, limit = 4, seed = 123)
```

---

rci

*Reliable change index*


---

## Description

**CAUTION! This function is still under development and not ready for use!** The `rciSC` function computes three indices of reliable change (Wise, 2004) and corresponding descriptive statistics.

## Usage

```
rci(data, dvar, pvar, rel, ci = 0.95, graph = FALSE, phases = c(1, 2))
```

## Arguments

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the <code>scdf</code> file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the <code>scdf</code> file.
<code>rel</code>	Reliability of the measure, used to compute the standard error.
<code>ci</code>	Width of confidence interval as a decimal. Default is <code>ci = 0.95</code> applying a 95%-confidence interval.
<code>graph</code>	If set <code>TRUE</code> , a box plot of phase A and B scores is displayed. <code>graph = FALSE</code> by default.
<code>phases</code>	A vector of two characters or numbers indicating the two phases that should be compared. E.g., <code>phases = c("A", "C")</code> or <code>phases = c(2, 4)</code> for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., <code>phases = list(A = c(1, 3), B = c(2, 4))</code> will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is <code>phases = c(1, 2)</code> .

## Author(s)

Juergen Wilbert

## References

Christensen, L., & Mendoza, J. L. (1986). A method of assessing change in a single subject: An alteration of the RC index. *Behavior Therapy, 17*, 305-308.

Hageman, W. J. J., & Arrindell, W. A. (1993). A further refinement of the reliable change (RC) index by improving the pre-post difference score: Introducing RCID. *Behaviour Research and Therapy, 31*, 693-700.

Jacobson, N. S., & Truax, P. (1991). Clinical Significance: A statistical approach to defining meaningful change in psychotherapy research. *Journal of Consulting and Clinical Psychology, 59*, 12-19.

Wise, E. A. (2004). Methods for analyzing psychotherapy outcomes: A review of clinical significance, reliable change, and recommendations for future directions. *Journal of Personality Assessment, 82*, 50 - 59.

## Examples

```
## Report the RCIs of the first case from the byHeart data and include a graph  
rci(byHeart2011[1], graph = TRUE, rel = 0.8)
```

---

read\_scdf

*Load single-case data from files*

---

## Description

Use the read\_scdf function to load single-case data csv, excel, or yaml files.

## Usage

```
read_scdf(  
  file,  
  cvar = "case",  
  pvar = "phase",  
  dvar = "values",  
  mvar = "mt",  
  sort_cases = FALSE,  
  phase_names = NULL,  
  type = NA,  
  sort.labels = NULL,  
  phase.names = NULL,  
  ...  
)
```

**Arguments**

file	Either a character string defining the file to be loaded (e.g. "SC_Anita.csv" (if left empty a dialog box for choosing will be opened) or a data.frame.
cvar	Sets the variable name of the "case" variable. Defaults to "case".
pvar	Sets the variable name of the "phase" variable. Defaults to "phase".
dvar	Sets the variable name of the "values" variable. Defaults to "values".
mvar	Sets the variable name of the "mt" variable. Defaults to "mt".
sort_cases, sort_labels	If set TRUE, the resulting list is sorted by label names (alphabetically increasing).
phase_names, phase.names	A character vector with phase names. Defaults to the phase names provided in the phase variable.
type	Format of the file to be loaded. Either "csv", "xlsx", "xls", "excel", "yaml" is possible. By default (NA) the type is extracted from the file extension.
...	Further arguments passed to the respective read function ( <a href="#">read.table</a> , <a href="#">read_excel</a> , <a href="#">read_yaml</a> command).

**Value**

Returns a single-case data frame. See [scdf](#) to learn about the format of these data frames.

**Author(s)**

Juergen Wilbert

**See Also**

[read.table](#), [readRDS](#), [read\\_excel](#)

Other io-functions: [convert\(\)](#), [write\\_scdf\(\)](#)

**Examples**

```
## Read SC-data from a file named "study1.csv" in your working directory
# study1 <- read_scdf("study1.csv")

## Read SC-data from a .csv-file with semicolon as field and comma as decimal separator
# study2 <- read_scdf("study2.csv", sep = ";", dec = ",")

## write_scdf and read_scdf
filename <- file.path(tempdir(), "test.csv")
write_scdf(exampleA1B1A2B2_zvt, filename)
dat <- read_scdf(filename, cvar = "case", pvar = "part", dvar = "zvt", mvar = "day")
res1 <- describe(exampleA1B1A2B2_zvt)$descriptives
res2 <- describe(dat)$descriptives
all.equal(res1, res2)
```

---

sample_names	<i>Samples random names</i>
--------------	-----------------------------

---

**Description**

Samples random names

**Usage**

```
sample_names(n = 1, type = "neutral", seed = NULL)
```

**Arguments**

n	Number of names
type	"neutral", "male", "female", or "mixed"
seed	A seed for the random number generator.

**Value**

A character vector with random names

**Examples**

```
sample_names(3)
```

---

scdf	<i>Single case data frame</i>
------	-------------------------------

---

**Description**

The scdf class stores single-case study data with one or more single-cases.

**Usage**

```
scdf(
  values,
  B_start,
  mt,
  phase,
  phase_design = NULL,
  name = NULL,
  dvar = "values",
  pvar = "phase",
  mvar = "mt",
  phase.design,
  B.start,
  ...
)
```

**Arguments**

<code>values</code>	A vector containing measurement values of the dependent variable.
<code>B_start</code> , <code>B.start</code>	The first measurement of phase B (simple coding if design is strictly AB).
<code>mt</code>	A vector defining measurement times. Default is <code>mt = (1, 2, 3, ..., n)</code> .
<code>phase</code>	A vector defining phase assignments.
<code>phase_design</code> , <code>phase.design</code>	A vector defining the length and label of each phase. E.g., <code>phase_design = c(A1 = 10, B1 = 10, A2 = 10, B2 = 10)</code> .
<code>name</code>	A name for the case.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
<code>mvar</code>	Character string with the name of the measurement time variable. Defaults to the attributes in the scdf file.
<code>...</code>	Additional variables. E.g., <code>teacher = c(0, 1, 0, 1, 0, 0, 1)</code> , <code>lesson = c(1, 3, 4, 5, 2, 3)</code> .

**Details**

If the dependent variable is a named vector then the names are extracted to create a phase design (e.g., `values = c(A = 2,3,5,4,3, B = 6,5,4,3)` will create an AB phase design with five and four measurements). An scdf contains several attributes: `dvar` The name of the dependent variable. `phase` The name of the phase variable. `mt` The name of the measurement time variable. `author` Information on the author of the data. `info` Further information on the data. E.g., a publication. `dvar`, `phase`, and `mt` are the defaults most of the scan function use. You can change the values of the attributes with the `scdf_attr` function (e.g., `scdf_attr(exampleAB_add, "dvar") <- "depression"` defines depression as the dependent variable. Please notice that all scan functions have arguments to define `dvar`, `phase`, and `mt` for a given analysis.

**Value**

Returns a single-case data frame `scdf` suitable for all functions of the scan package. Multiple data sets (e.g. from Multiple Baseline Designs) can be listed.

**Author(s)**

Juergen Wilbert

**See Also**

Other data manipulation functions: [add\\_l2\(\)](#), [as.data.frame.scdf\(\)](#), [as\\_scdf\(\)](#), [fill\\_missing\(\)](#), [moving\\_median\(\)](#), [outlier\(\)](#), [ranks\(\)](#), [select\\_cases\(\)](#), [set\\_vars\(\)](#), [shift\(\)](#), [smooth\\_cases\(\)](#), [standardize\(\)](#), [truncate\\_phase\(\)](#)

**Examples**

```

## Scores on a letter naming task were collected on eleven days in a row.
## The intervention started after the fifth measurement,
## so the first B phase measurement was 6 (B_start = 6).
klaas <- scdf(
  c(5, 7, 8, 5, 7, 12, 16, 18, 15, 14, 19),
  B_start = 6, name = "Klaas"
)
plot(klaas)

# Alternative coding 1:
klaas <- scdf(
  c(A = 5, 7, 8, 5, 7, B = 12, 16, 18, 15, 14, 19),
  name = "Klaas"
)

# Alternative coding 2:
klaas <- scdf(
  c(5, 7, 8, 5, 7, 12, 16, 18, 15, 14, 19),
  phase_design = c(A = 5, B = 6), name = "Klaas"
)

## Unfortunately in a similar study there were no data collected on
## days 3 and 9. Use NA to pass them to the function:
emmi <- scdf(c(5, 7, NA, 5, 7, 12, 16, 18, NA, 14, 19),
  phase_design = c(A = 5, B = 6), name = "Emmi"
)
describe(emmi)

## In a MBD over three cases, data were collected eleven days in a row.
## Intervention starting points differ between subjects as they were
## randomly assigned. The three SCDFs are then combined in a list for
## further conjoined analyses.
charlotte <- scdf(c(A = 5, 7, 10, 5, 12, B = 7, 10, 18, 15, 14, 19))
theresa <- scdf(c(A = 3, 4, 3, 5, B = 7, 4, 7, 9, 8, 10, 12))
antonia <- scdf(c(A = 9, 8, 8, 7, 5, 7, B = 6, 14, 15, 12, 16))
mbd <- c(charlotte, theresa, antonia)
names(mbd) <- c("Charlotte", "Theresa", "Antonia")
overlap(mbd)

## In a classroom-based intervention it was not possible to measure outcomes
## every day, but only on schooldays. The sequence of measurements is passed
## to the package by using a vector of measurement times.
frida <- scdf(
  c(A = 3, 2, 4, 2, 2, 3, 5, 6, B = 8, 10, 8, 12, 14, 13, 12),
  mt = c(1, 2, 3, 4, 5, 8, 9, 10, 11, 12, 14, 15, 16, 17, 18)
)
summary(frída)
describe(frída)

## example with two independent variables and four phases

```

```
jim <- scdf(
  zvt = c(47, 58, 76, 63, 71, 59, 64, 69, 72, 77, 76, 73),
  d2 = c(131, 134, 141, 141, 140, 140, 138, 140, 141, 140, 138, 140),
  phase_design = c(A1 = 3, B1 = 3, A2 = 3, B2 = 3), dvar = "zvt"
)
overlap(jim, phases = list(c("A1", "A2"), c("B1", "B2")))
```

---

 select\_cases

*Select a subset of cases*


---

## Description

Select a subset of cases

## Usage

```
select_cases(scdf, ...)
```

## Arguments

scdf	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
...	Selection criteria. Either numeric, objectnames, or as characters.

## Value

An scdf with a subset of cases

## See Also

Other data manipulation functions: [add\\_l2\(\)](#), [as.data.frame.scdf\(\)](#), [as\\_scdf\(\)](#), [fill\\_missing\(\)](#), [moving\\_median\(\)](#), [outlier\(\)](#), [ranks\(\)](#), [scdf](#), [set\\_vars\(\)](#), [shift\(\)](#), [smooth\\_cases\(\)](#), [standardize\(\)](#), [truncate\\_phase\(\)](#)

## Examples

```
select_cases(exampleAB, Johanna, Karolina)
select_cases(exampleAB, c(Johanna, Karolina))
select_cases(exampleAB, 1,2)
select_cases(exampleAB, 1:2)
select_cases(exampleAB, -Johanna)
select_cases(exampleAB, -c(Johanna, Karolina))
v <- c("Moritz", "Jannis")
select_cases(exampleA1B1A2B2, v)
```

---

select_phases	<i>Select and combine phases for overlap analyses</i>
---------------	-------------------------------------------------------

---

**Description**

Useful when working with

**Usage**

```
select_phases(data, A, B)
```

**Arguments**

data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
A	Selection of the A phase
B	Selection of the B phase

**Value**

An scdf with selected phases

**Examples**

```
exampleA1B1A2B2_zvt %>%  
  select_phases(A = c(1, 3), B = c(2, 4)) %>%  
  overlap()
```

---

set_vars	<i>Set analysis variables in an scdf</i>
----------	------------------------------------------

---

**Description**

Set analysis variables in an scdf

**Usage**

```
set_vars(data, dvar, mvar, pvar)  
  
set_dvar(data, dvar)  
  
set_mvar(data, mvar)  
  
set_pvar(data, pvar)
```

**Arguments**

data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
dvar	Character string. Name of the dependent variable.
mvar	Character string. Name of the measurement-time variable.
pvar	Character string. Name of the phase variable.

**See Also**

Other data manipulation functions: [add\\_l2\(\)](#), [as.data.frame.scdf\(\)](#), [as\\_scdf\(\)](#), [fill\\_missing\(\)](#), [moving\\_median\(\)](#), [outlier\(\)](#), [ranks\(\)](#), [scdf](#), [select\\_cases\(\)](#), [shift\(\)](#), [smooth\\_cases\(\)](#), [standardize\(\)](#), [truncate\\_phase\(\)](#)

**Examples**

```
exampleAB_add %>%
  set_dvar("depression") %>%
  describe()
```

---

show	<i>Show scdf cases in viewer</i>
------	----------------------------------

---

**Description**

Each case with in scdf opens a separate viewer tab.

**Usage**

```
show(scdf, max = 5)
```

**Arguments**

scdf	A scdf object
max	Maximum number of cases opened

**Examples**

```
## show(exampleABC)
```

---

smd *Standardized mean differences*

---

**Description**

The `smd` function provides various standardized mean effect sizes for single-case data.

**Usage**

```
smd(data, dvar, pvar, mvar, decreasing = FALSE, phases = c(1, 2))
```

**Arguments**

<code>data</code>	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
<code>dvar</code>	Character string with the name of the dependent variable. Defaults to the attributes in the <code>scdf</code> file.
<code>pvar</code>	Character string with the name of the phase variable. Defaults to the attributes in the <code>scdf</code> file.
<code>mvar</code>	Character string with the name of the measurement time variable. Defaults to the attributes in the <code>scdf</code> file.
<code>decreasing</code>	If you expect data to be lower in the B phase, set <code>decreasing = TRUE</code> . Default is <code>decreasing = FALSE</code> .
<code>phases</code>	A vector of two characters or numbers indicating the two phases that should be compared. E.g., <code>phases = c("A", "C")</code> or <code>phases = c(2, 4)</code> for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., <code>phases = list(A = c(1, 3), B = c(2, 4))</code> will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is <code>phases = c(1, 2)</code> .

**Details**

'sd cohen' is the (unweighted) average of the variance of phase A and B. 'sd Hedges' is the weighted average of the variance of phase A and B (with a degrees of freedom correction). 'Hedges' g' is the mean difference divided by 'sd Hedges'. 'Hedges' g correction' and 'Hedges' g durlak correction' are two approaches of correcting Hedges' g for small sample sizes. 'Glass' delta' is the mean difference divided by the standard deviation of the A-phase. 'Cohens d' is the mean difference divided by 'sd cohen'.

**Author(s)**

Juergen Wilbert

**See Also**

[overlap](#), [describe](#)

**Examples**

```
smd(exampleAB)
```

---

```
style_plot
```

---

*Create styles for single-case data plots*

---

**Description**

The `style_plot` function is used to create graphical styles for a single-case plot

**Usage**

```
style_plot(style = "default", ...)
```

**Arguments**

<code>style</code>	A character string or a vector of character strings with predefined styles.
<code>...</code>	Further arguments passed to the plot command.

**Details**

`style_plot("")` will return a list of predefined styles. Predefined styles can be combined `style_plot(style = c("grid2", "tiny"))` where settings of a latter style overwrite settings of the former. Additional style parameters are set following the style argument and can be combined with those: `style_plot(style = "grid2", fill = "grey50", pch = 18)`.

**Value**

Returns a list to be provided for the style argument of the `plot.scdf` function.

- `fill` If set, the area under the line is filled with the given color (e.g., `fill = "tomato"`). Use the standard R command `colors()` to get a list of all possible colours. `fill` is empty by default.
- `annotations` A list of parameters defining annotations to each data point. This adds the score of each MT to your plot.
  - `"pos"` Position of the annotations: 1 = below, 2 = left, 3 = above, 4 = right.
  - `"col"` Color of the annotations.
  - `"cex"` Size of the annotations.
  - `"round"` Rounds the values to the specified decimal.
- `annotations = list(pos = 3, col = "brown", round = 1)` adds scores rounded to one decimal above the data point in brown color to the plot.
- `"names"` A list of parameters defining the depiction of phase names (e.g. `names = list(cex = 0.8, col = "red", side = 1)`: `cex` for size, `col` for color, and `side` for position). See `mttext` for more details.
- `"lwd"` Width of the plot line. Default is `lwd = 2`.

- "pch" Point type. Default is pch = 17 (triangles). Other options are for example: 16 (filled circles) or "A" (uses the letter A).
- "main" Main title of the plot.
- "mai" Sets the margins of the plot.
- "bty" Shape of the frame surrounding the inner plot
- "fill.bg" Background color of the plot. If a vector is provided, these colors will be assigned to phases (each phase name becomes a color).
- "grid" Color of a grid.
- "text.ABlag" Text displayed between phases.
- "cex.axis" Size of the axis annotations
- "las" Orientation of the axis annotations
- "col.lines" Color of the lines
- "col.dots" Color of the dots
- "col.seperator" Color of the phase seperating lines
- "col.bg" Color of the outer plot
- "col" General color setting for the plot
- "col.text" Color of all labels of the plot.

### Author(s)

Juergen Wilbert

### See Also

[plot.scdf](#)

### Examples

```
newstyle <- style_plot(style = "default")
newstyle$text.ABlag <- c("START", "END")
newstyle$col.dots <- ""
newstyle$annotations <- list(cex = 0.6, col = "grey10", offset = 0.4)
newstyle$names <- list(cex = 0.8, col = "blue", side = 1, adj = 1, line = -1, at = 31)
newstyle$fill.bg <- c("grey99", "grey95", "grey90")
plot(exampleABC, style = newstyle, main = "Example Plot")
```

---

subset.scdf	<i>Subset cases, rows, and variables</i>
-------------	------------------------------------------

---

**Description**

Subset cases, rows, and variables

**Usage**

```
## S3 method for class 'scdf'
subset(x, filter, select, cases, ...)
```

**Arguments**

x	An scdf object.
filter	Logical expression indicating rows to keep: missing values are taken as false.
select	Expression, indicating columns to select from an scdf.
cases	Expression, indicating cases to keep from an scdf.
...	[not implemented]

**Value**

An scdf.

**Examples**

```
subset(exampleAB, (values < 60 & phase == "A") | (values >= 60 & phase == "B"))
subset(exampleAB_add, select = c(-cigarrets, -depression))
subset(exampleAB, cases = c(Karolina, Johanna))
subset(exampleA1B1A2B2, phase %in% c("A1", "B2"), cases = Pawel:Moritz)
```

---

summary.scdf	<i>Summary function for an scdf</i>
--------------	-------------------------------------

---

**Description**

Summary function for an scdf

**Usage**

```
## S3 method for class 'scdf'
summary(object, all_cases = FALSE, ...)
```

**Arguments**

object	scdf
all_cases	IF TRUE, more that 10 cases are summarized
...	not in use

---

tau_u	<i>Tau-U for single-case data</i>
-------	-----------------------------------

---

**Description**

This function calculates indices of the Tau-U family as proposed by Parker et al. (2011a).

**Usage**

```
tau_u(
  data,
  dvar,
  pvar,
  tau_method = c("b", "a"),
  method = c("complete", "parker"),
  phases = c(1, 2),
  meta_analyses = TRUE,
  ci = 0.95,
  ci_method = c("z", "tau"),
  meta_weight_method = c("z", "tau"),
  continuity_correction = FALSE,
  meta_method = NULL
)
```

**Arguments**

data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
dvar	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
pvar	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
tau_method	Character with values "a" or "b" (default) indicating whether Kendall Tau A or Kendall Tau B is applied.
method	"complete" (default) or "parker". The latter calculates the number of possible pairs as described in Parker et al. (2011) which might lead to tau-U values greater than 1.
phases	A vector of two characters or numbers indicating the two phases that should be compared. E.g., phases = c("A", "C") or phases = c(2, 4) for comparing the second to the fourth phase. Phases could be combined by providing a list with two elements. E.g., phases = list(A = c(1, 3), B = c(2, 4)) will compare phases 1 and 3 (as A) against 2 and 4 (as B). Default is phases = c(1, 2).

meta_analyses	If TRUE, a meta analysis is conducted.
ci	Confidence intervals
ci_method	String to specify the method for calculating the standard error of tau. Either "tau", "z", or "s" (not recommended).
meta_weight_method	String to specify the method for calculating the weights of the studies. Either "tau" or "z".
continuity_correction	If TRUE, a continuity correction is applied for calculating p-values of correlations (here: S will be reduced by one before calculating Z)
meta_method	(not implemented) All meta analyses are based on a fixed model.

### Details

Tau-U is an inconsistently operationalized construct. Parker et al. (2011b) describe a method which may result in Tau-U larger than 1. A different implementation of the method (provided at <http://www.singlecaseresearch.org/calculators/tau-u>) uses tau-b (instead of tau-a as in the original formulation by Parker). Bossart et. al (2018) describe inconsistencies in the results from this implementation as well. Another problems lies in the calculation in overall Tau-U values from several single cases. The function presented here applies a metaanalyzes to gain the overall values. Each tau value is weighted by the inverse of the variance (ie. the tau standard error). The confidence intervals for single cases are calculated by Fisher-Z transforming tau, calculating the confidence intervals, and inverse transform them back to tau (see Long & Cliff, 1997).

### Value

table	A data frame containing statistics from the Tau-U family, including: Pairs, positive and negative comparisons, S, and Tau
matrix	The matrix of comparisons used for calculating the statistics.
tau_u	Tau-U value.

### Author(s)

Juergen Wilbert

### References

- Brossart, D. F., Laird, V. C., & Armstrong, T. W. (2018). Interpreting Kendall's Tau and Tau-U for single-case experimental designs. *Cogent Psychology*, 5(1), 1–26. <https://doi.org/10.1080/23311908.2018.1518687>.
- Long, J. D., & Cliff, N. (1997). Confidence intervals for Kendall's tau. *British Journal of Mathematical and Statistical Psychology*, 50(1), 31–41. <https://doi.org/10.1111/j.2044-8317.1997.tb01100.x>
- Parker, R. I., Vannest, K. J., & Davis, J. L. (2011a). Effect Size in Single-Case Research: A Review of Nine Nonoverlap Techniques. *Behavior Modification*, 35(4), 303–322. <https://doi.org/10/dsdfs4>
- Parker, R. I., Vannest, K. J., Davis, J. L., & Sauber, S. B. (2011b). Combining Nonoverlap and Trend for Single-Case Research: Tau-U. *Behavior Therapy*, 42, 284-299.

**See Also**

Other overlap functions: [corrected\\_tau\(\)](#), [nap\(\)](#), [overlap\(\)](#), [pand\(\)](#), [pem\(\)](#), [pet\(\)](#), [pnd\(\)](#)

**Examples**

```
tau_u(Grosche2011$Eva)

## Replicate tau-U calculation from Parker et al. (2011)
bob <- scdf(c(A = 2, 3, 5, 3, B = 4, 5, 5, 7, 6), name = "Bob")
res <- tau_u(bob, method = "parker", tau_method = "a")
print(res, complete = TRUE)

## Request tau-U for all single-cases from the Grosche2011 data set
tau_u(Grosche2011)
```

---

trend

*Trend analysis for single-cases data*


---

**Description**

The trend function provides an overview of linear trends in single case data. By default, it provides the intercept and slope of a linear and quadratic regression of measurement time on scores. Models are calculated separately for each phase and across all phases. For more advanced use, you can add regression models using the R-specific formula class.

**Usage**

```
trend(
  data,
  dvar,
  pvar,
  mvar,
  offset = "deprecated",
  first_mt = 0,
  model = NULL
)
```

**Arguments**

data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
dvar	Character string with the name of the dependent variable. Defaults to the attributes in the scdf file.
pvar	Character string with the name of the phase variable. Defaults to the attributes in the scdf file.
mvar	Character string with the name of the measurement time variable. Defaults to the attributes in the scdf file.

offset	(Deprecated. Please use first_mt). An offset for the first measurement-time of each phase. If offset = 0, the phase measurement is handled as MT 1. Default is offset = -1, setting the first value of MT to 0.
first_mt	A numeric setting the value for the first measurement-time. Default = 0.
model	A string or a list of (named) strings each depicting one regression model. This is a formula expression of the standard R class. The parameters of the model are values, mt and phase.

**Value**

trend	A matrix containing the results (Intercept, B and beta) of separate regression models for phase A, phase B, and the whole data.
offset	Numeric argument from function call (see Arguments section).

**Author(s)**

Juergen Wilbert

**See Also**

[describe](#)

Other regression functions: [autocorr\(\)](#), [corrected\\_tau\(\)](#), [hplm\(\)](#), [mplm\(\)](#), [plm\(\)](#)

**Examples**

```
## Compute the linear and squared regression for a random single-case
design <- design(slope = 0.5)
matthea <- random_scdf(design)
trend(matthea)

## Besides the linear and squared regression models compute two custom models:
## a) a cubic model, and b) the values predicted by the natural logarithm of the
## measurement time.
design <- design(slope = 0.3)
ben <- random_scdf(design)
trend(ben, offset = 0, model = c("Cubic" = values ~ I(mt^3), "Log Time" = values ~ log(mt)))
```

---

write\_scdf

*Data output*

---

**Description**

This function restructures and writes single-case data into a .csv-file.

**Usage**

```
write_scdf(data, filename = NULL, sep = ",", dec = ".", ...)
```

**Arguments**

data	A single-case data frame. See <a href="#">scdf</a> to learn about this format.
filename	A character string defining the output file name (e.g. "SC_data.csv").
sep	The field separator string. Values within each row of x are separated by this string.
dec	The string to use for decimal points in numeric or complex columns: must be a single character.
...	Further arguments passed to write.table.

**Details**

This is a wrapper for the write.table function with predefined parameters.

**Author(s)**

Juergen Wilbert

**See Also**

[write.table](#), [saveRDS](#)

Other io-functions: [convert\(\)](#), [read\\_scdf\(\)](#)

**Examples**

```
## write single-case data to a .csv-file
filename <- tempfile(fileext = ".csv")
jessica <- random_scdf(design(level = .5))
write_scdf(jessica, tempfile())

## write multiple cases to a .csv-file with semicolon as field and comma as
## decimal separator
write_scdf(Grosche2011, filename, sep = ";", dec = ",")

## read_scdf and write_scdf
write_scdf(exampleA1B1A2B2_zvt, filename)
dat <- read_scdf(filename, cvar = "case", pvar = "part",
                dvar = "zvt", mvar = "day")
res1 <- describe(exampleA1B1A2B2_zvt)$descriptives
res2 <- describe(dat)$descriptives
all.equal(res1, res2)
```

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